# © 2012 IJFANS. All Rights Reserved, UGC CARE Listed (Group -1) Journal Volume 11, Iss 1, 2022 Analysing the concentration of phycobilin pigment in nostro mussel at different pH levels

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### Abstract:

Nostoc muscorum is a free-living microbe that can be found all over the world in a wide variety of terrestrial and aquatic settings. It is popular in desert crusts and benthic communities of water-logged paddy fields and is known to create symbiotic partnerships with terrestrial plants. N. muscorum prefers environments with pH values between 7.0 and 8.5. It was cultured in a medium with a pH range of 7 to 11, and its morphological and physiological traits, such as the frequency of heterocysts and the amount of the pigment phycobilin, were assessed. The findings show that N. muscorum can develop and carry out its physiological functions at an alkaline pH range of 8 to 10.

Key Words: Ph, Phycobilin, Heterocyst,

# **INTRODUCTION**

Cyanobacteria can be found in almost every terrestrial and aquatic habitat oceans, fresh water, damp soil, temporarily moistened rocks in deserts, bare rock and soil, and even Antarctic rocks. They can occur as planktonic cells or form phototrophic biofilms. They are found in almost every endolithic ecosystem (Grube, M. et. al., 2007). A few are endosymbionts in lichens, plants, various protists, or sponges and provide energy for the host (Vaughan et al., 2011). Cyanobacteria fulfill vital ecological functions in the world's oceans, being important contributors to global carbon and nitrogen budgets.Cyanobacteria also form symbiotic association with animals and plants. Symbiotic relations exist with, for example, fungi, bryophytes, pteridophytes, gymnosperms and angiosperms (Rai, 2002). *Nostocmuscorum* is a

© 2012 IJFANS. All Rights Reserved, UGC CARE Listed (Group -1) Journal Volume 11, Iss 1, 2022 *Research paper* free-living microorganism is distributed over a large area of the globe in many varying habitats, both aquatic and terrestrial. Its ability to populate such a diverse range of conditions is due the cellular development of metabolic adaptations to harsh conditions, such as desiccation-tolerance, salt-tolerance, and nitrogen-fixation processes. As cyanobacteria are phototrophic, performing photosynthesis in their environments requiring  $CO_2$  for growth and also fixing atmospheric nitrogen. The ideal environment for *N. muscorum* is one with pH in the range of 7.0 to 8.5, with a lower pH limit of 5.7 (Allison et al., 1937). It grows best when light intensity is less than that of direct sunlight, (Blumwald& Tel-Or, 1982). *N. muscorum* has heterocyst, which are specialized nitrogen-fixation cells. Heterocyst, (5-10% of cells) appear when it is transferred to nitrogen free media. Appearance of heterocyst is concurrent with an increase in nitrogenase activity, which reduces N<sub>2</sub> to NH<sub>3</sub>. It fixes nitrogen that is important in symbiotic relationships with fungi, liverworts, hornworts, mosses, cycads (Dodds and Gudder, 1995).

# MATERIALS AND METHODS

# 1. **Establishment of culture**:

The algal culture was established in liquid Fogg's medium (0.2gm MgSO<sub>4</sub>.7H<sub>2</sub>O, 0.2gm K<sub>2</sub>HPO<sub>4</sub>, 0.1gm CaCl<sub>2</sub>.H<sub>2</sub>O, 5 ml FeEDTA (dissolve 0.745gm Na<sub>2</sub>EDTA in hot water & add 0.557gm FeSO<sub>4</sub>, boil to dissolve completely, final volume to 100ml), 1ml/litre, micronutrients (286 mg H<sub>3</sub>BO<sub>3</sub>,18 mg MnCl<sub>2</sub>.4H<sub>2</sub>O, 22 mg ZnSO<sub>4</sub>.7H<sub>2</sub>O, 39 mg Na<sub>2</sub>MoO<sub>4</sub>.2H<sub>2</sub>O, 8mg CuSO<sub>4</sub>, 4 mg CoCl<sub>2</sub>), pH 7.5, final volume to 1 liter.)

5g of algal inoculums was added to each culture bottle in aseptic conditions. The culture was maintained in the diffused sun light for the period 30 days to obtain sufficient growth. Alga was sub-cultured in open culture system using culture trays. 1 liter of Fogg's medium was added per tray. The culture trays were divided into eight sets of different pH from pH 4 to pH 11 each containing 15 trays. This culture was allow to grow for 1 week & then used for further analysis.

# 2. Measurement of pH:

The initial pH was measured after 1 week of subculture. The change in pH was observed. The Fogg's medium of pH 7.5 was added to each set and then pH was measured on each day for successive days.

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#### Research paper 3. Study of Morphology:

The morphological observation with respect to growth and thallus morphology was done for each set of pH and the observations were recorded.

# 4. Determination of Heterocyst Frequency:

The slides were prepared for each set of pH and observed under compound microscope (45x magnification). Number of heterocyst were counted from different 5 fields on slide and mean was calculated and frequency was determined for each set.

# 5. Estimation of PhycobilinPigment Content:

The total phycobilin pigment content was determined by protocol given by Evans (1988).0.5g of algal culture was crushed in the 5 ml of 0.1M phosphate buffer, pH 6.8(25.5 ml 0.1M NaH2PO4, 24.5 ml Na2HPO4, final volume 100 ml, pH 6.8).with acid washed sand using chilled mortar& pestle. This homogenate was transferred into centrifuge tube & spin at 5000 rpm for 10 min. Final volume of supernatant was set to 25 ml using 0.1M phosphate buffer, pH 6.8. The resultant solution was used as sample for further investigation. The absorbance was measured using spectrophotometer at wavelength 455nm, 564nm, 592nm, 618nm, 645nm and 650nm; for c-Phycocyanin (c-PC), c-Phycoerythrin (c-PE) and Allophycocyanin (APC) Amount of c-PC, c-PE and APC was determined by using formulae as given in protocol.

# **RESULTS AND DISCUSSION**

The morphological study reveals that growth of *Nostocmuscorum* at acidic pH from pH 4-pH 5 was less as compared to neutral and alkaline pH ranging from pH 8-pH 11. The color of thalii observed was from yellowish green to dark green with increase in pH. (Figure No.1 &2, Table No. 1)



Figure No. 1: Culture of *N. muscorum* 

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Figure No. 2: Sub-culture of N. muscorum

pН	Morphological Features
4	Less growth, yellowish green
5	Less growth, yellowish green
6	Moderate growth, thick, dark green
7	Moderate growth, thick, light green
8	High growth, pale yellowish green
9	High growth, thick, greenish yellow
10	Highest growth rate, dark green
11	Highest growth rate, dark green

# Table No. 1: Morphology

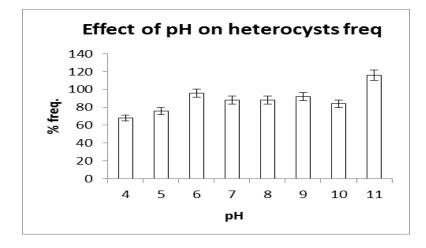
Table No.	2:	Heterocyst	Frequency
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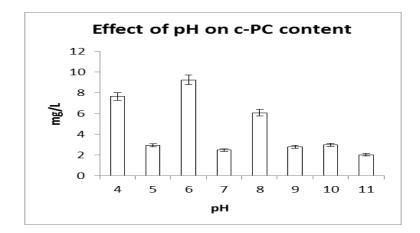
pН	Mean Heterocyst	Frequency %
4	3.4	68
5	3.8	76
6	4.8	96
7	4.4	88
8	4.4	88
9	4.6	92
10	4.2	84
11	5.8	116

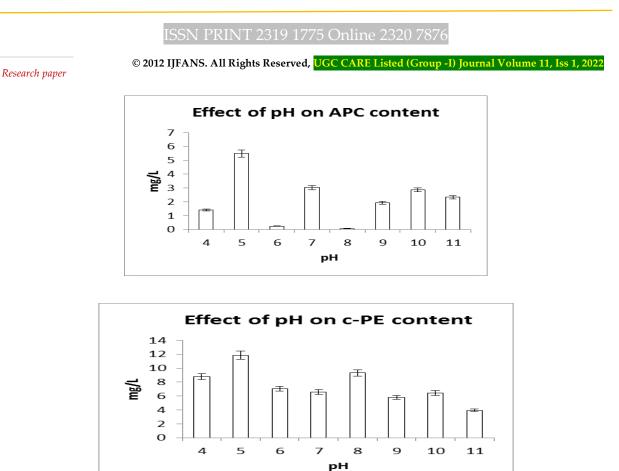
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Research paper **Table No. 3:** c-Phycocyanin, Allophycocyanin and c-Phycoerythrin Content

pН	c-PC (mg/L)	APC (mg/L)	c-PE (mg/L)
4	7.642	1.4097	8.8014
5	2.9584	5.4911	11.8714
6	9.228	0.2518	7.0363
7	2.496	3.031	6.5688
8	6.0744	0.07788	9.3278
9	2.812	1.9356	5.8121
10	2.994	2.8741	6.4344
11	2.04	2.3448	3.9686







According to earlier studies, it is found that cyanobacteria cultured at three different pH conditions as, 7.0, 9.5, 10.5. The higher value of growth rate, cell yield observed in pH 10.5. The low pH limits the growth in terms of no. of cells & biomass. (Figure No.2, Table No. 1)

The heterocyst frequency was found to be increased with increase in pH 4 to pH 11. This concludes that maximum no. of heterocyst are formed in alkaline pH than the acidic pH. (Table No. 3)

Among the studied phycobilin pigments content was observed at acidic pH 5 and minimum amount in alkaline pH 8. The same trend was observed for the carotenoid content also. (Table No. 4)

# CONCLUSION

Effect of pH on morphological and physiological characteristics of Nostocmuscorum:

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Research paper The data on growth, number of heterocyst, chlorophyll pigment, carotenoid that the organism has ability to create and maintain the favorable growth conditions for its growth by temporary changes in metabolism.

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