Research paper

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An Overview of Microbial Production of Lactic Acid from Molasses

^aArun Kumar Mahato, ^b Leelawati Kumari

^{a,b,} University, PG, Department of Chemistry, Binod Bihari Mahto Koyalanchal University, Dhanbad, Jharkhand, India

Corresponding author

1. Arun Kumar Mahato,

University, PG, Department of Chemistry, Binod Bihari Mahato Koyalanchal University, Dhanbad, Jharkhand, India

shakticollegesijua78@gmail.com

ABSTRACT: Lactic acid is a versatile organic acid with a wide range of industrial applications, spanning from the food and pharmaceutical sectors to biodegradable plastics and environmental remediation. This review paper aims to provide detailed information about lactic acid, encompassing its production, properties, and applications. The microbial production of lactic acid from molasses offers a promising and eco-friendly alternative to traditional chemical synthesis methods. As technology and bioprocesses continue to advance, this approach is poised to become increasingly competitive and sustainable, effectively addressing the growing demand for lactic acid across various industries. In recent years, microbial lactic acid production has garnered significant attention due to its sustainability and cost-effectiveness. Molasses, a byproduct of the sugar industry, serves as an abundant and cost-effective substrate for lactic acid production. This overview also delves into key aspects of microbial lactic acid production from molasses, including the selection of microorganisms, fermentation processes, optimization strategies, and downstream processing. It highlights both the advantages and



challenges associated with this biotechnological approach, emphasizing its potential as a sustainable and eco-friendly pathway for lactic acid production.

This study underscores the growing significance of microbial lactic acid production from molasses as a promising alternative to traditional chemical synthesis, with wide-reaching implications for various industries and environmental sustainability. Additionally, this paper explores the physical and microbial production of lactic acid from sucrose-derived raw materials, with molasses emerging as the most cost-effective choice

KEYWORDS: Lactic acid, Lactic, Molasses, microbial production, Cheap

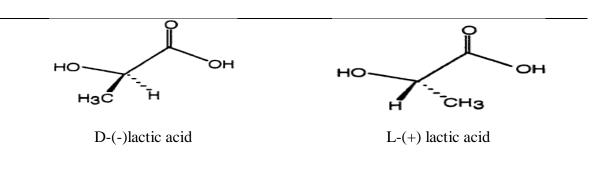
1. Introduction

Lactic acid is a natural organic acid with a wide array of applications in the pharmaceutical, chemical, food, and healthcare industries and holds significant importance in pharmaceutical preparations, electroplating, the leather industry, and the pork industry, production of polypropylene oxide and biodegradable polylactic acid ¹⁻⁴. The fermentation process ensures the rapid and reliable generation of lactic acid, representing one of the earliest microbial products in history⁵. Recent years have witnessed a surge in interest in lactic acid production from biomass through fermentation process which brings garners attention due to its advantages over chemical synthesis ⁶⁻⁸. In recent years, much work has been carried out on the optimization of lactic acid production from different biomass sources⁹⁻¹⁴. Lactic acid is an organic acid. It is 2-hydroxypropionic acid and occurs in two isomeric forms, D or L, namely, (L-(+)-LA and D-(-)-LA) as shown below



IJFANS INTERNATIONAL JOURNAL OF FOOD AND NUTRITIONAL SCIENCES ISSN PRINT 2319 1775 Online 2320 7876

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2. **Properties of Lactic acid**: Physical and chemical properties of lactic acid have been incorporated in Table1.

Properties of Lactic acid¹⁵⁻²²

Property	Value or Description
Formula	CH3CH(OH)COOH
Molecular weight	90.08 g/mol
Odour	Odourless
Odor Threshold	The odor threshold for lactic acid is low, and it can be detected at low concentrations.
Taste	Mild acid taste
Toxicity	Oral rat LD50. 3543mg/kg
Synonyms	2-hydroxypropanoic acid,
	1-hydroxyethanecarboxylic acid
	Ethylidenelactic acid,
	Alph hydroxypropionic acid
Physical state	Colourless to slightly yellow, syrupy liquid
Melting point	17°C
Boiling point	122 °C
Specific gravity	1.2
Solubility in water	Miscible
NFPA rating	Health3, Flamability 1, Reactivity 1
Flash point	112 °C
Stability	Stable under ordinary conditions
K _a	1.38×10^{-4}
pK _a	Lactic acid has two pKa values: 3.86 and 5.12, reflecting its two ionizable hydrogen atoms. The pKa values can change depending on concentration
Appearance	Colorless or slightly yellow liquid



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State at Room Temperature	Liquid			
Density	1.209 g/cm ³ (at 20 °C)			
Solubility	Soluble in water, ethanol, and ether			
Hygroscopicity	Absorbs moisture from the air			
pH	2.4 (for 1 M solution)			
Specific Gravity	1.21 - 1.22 (at 20 °C)			
Viscosity	1.21 cP (at 25 °C)			
Heat of Vaporization	42.8 kJ/mol			
Freezing Point	Approximately 53 °C (127 °F) (for 80% solution)			
Autoignition Temperature	Not applicable (lactic acid does not autoignite)			
Electrical Conductivity	Lactic acid is a weak electrolyte and has low electrical conductivity in its pure form.			
Color	Lactic acid is typically colorless, but it may appear slightly yellow in impure or concentrated forms.			
Heat of Formation	Approximately -694 kJ/mol			
Heat Capacity (Cp)	Approximately 2.62 J/(g·K) at 25 °C			
Critical Temperature	Approximately 242 °C (467.6 °F)			
Partition Coefficient (log P)	Around -0.57 (n-octanol/water)			
Surface Tension	Approximately 49 mN/m (at 25 °C)			
Solubility in Organic Solvents	Lactic acid is soluble in various organic solvents such as ethanol, acetone, and ethyl acetate.			
Heat of Solution	-55 kJ/mol (when dissolved in water)			
Vapor Pressure (at 20 °C)	Very low, typically less than 1 mmHg			
Octanol-Water Partition Coefficient (log P)	Approximately -0.67			
Optical Activity	Lactic acid exists in two optical isomers, L- lactic acid and D-lactic acid, each with different optical activity. The optical rotation depends on the isomer and its concentration.			
Specific Heat Capacity	Approximately 2.44 J/g°C (for L-lactic acid)			
Combustibility	Lactic acid is not considered highly flammable, but it can support combustion under certain conditions.			
Refractive Index	Approximately 1.42 (for pure lactic acid)			
Solubility in Ether	Lactic acid is soluble in ether.			
Solubility in Acetone	Lactic acid is soluble in acetone.			
Surface Density	Approximately 7.2 mg/cm^2			
Solubility in Ethanol	Lactic acid is soluble in ethanol.			



Optical Rotation	The optical rotation of lactic acid varies based on the specific optical isomer (L-lactic acid or D-lactic acid) and its concentration. It exhibits optical activity.			
Standard Enthalpy of Formation	Approximately -694.6 kJ/mol			
Partition Coefficient (log Kow)	Approximately -1.02 (for n-octanol/water)			
Molecular Geometry	Lactic acid has a pyramidal geometry around the carbon atom.			
Standard Gibbs Free Energy of Formation	Approximately -564.70 kJ/mol (at 298.15 K and 1 bar)			
Magnetic Susceptibility	Lactic acid is diamagnetic, meaning it is not attracted to a magnetic field.			
Specific Conductance	Lactic acid solutions exhibit low electrical conductivity and are weak electrolytes.			
Environmental Fate	Lactic acid is biodegradable and considered environmentally friendly. It is typically broken down by microorganisms in natural systems.			
Chirality	Lactic acid has chirality and exists in two optical isomers, L-lactic acid, and D-lactic acid, each with distinct optical properties.			
Optical Purity	Lactic acid enantiomers may be used to express optical purity, often denoted as %ee (percent enantiomeric excess).			
Acidity	Lactic acid is a weak acid with a pKa value of approximately 3.86 for the carboxylic acid group.			
Partition Coefficient (log P)	Approximately -0.67 (for n-octanol/water)			
Refractive Index	Approximately 1.42 (for pure lactic acid)			
Heat of Solution	Approximately -55 kJ/mol (when dissolved in water)			
Critical Density	0.22 g/cm ³			
Heat of Fusion	10.3 kJ/mol			
Heat of Combustion	-1,265 kJ/mol (when burned)			
Partition Coefficient (log Kow)	-1.02 (for n-octanol/water)			
Molecular Geometry	Lactic acid has a pyramidal geometry around the carbon atom.			
Standard Gibbs Free Energy of Formation	-564.70 kJ/mol (at 298.15 K and 1 bar)			
Electrical Conductivity	Lactic acid is a weak electrolyte and has low electrical conductivity in its pure form.			
Specific Heat Capacity (Cp)	2.42 J/(g·K) at 25 °C			
Density (at 25°C)	1.206 g/cm ³			



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Refractive Index (at 20°C)	1.42 (for pure lactic acid)		
Solubility in Organic Solvents	Lactic acid is soluble in various organic solvents, such as acetone, ethyl acetate, and ethanol.		
Vapor Density	Lactic acid vapor is heavier than air.		
Partition Coefficient (log P)	Approximately -0.67 (for n-octanol/water).		
Environmental Impact	Lactic acid is biodegradable and considered environmentally friendly as it can be broken down by microorganisms in natural systems.		
Critical Pressure	Approximately 4.8 MPa (48 bar)		
Heat of Solution	Approximately -55 kJ/mol (when dissolved in water)		
Optical Rotation	The optical rotation of lactic acid varies based on the specific optical isomer (L-lactic acid or D-lactic acid) and its concentration. It exhibits optical activity.		

- **3.** Carbon sources : Lactic acid can be produced through fermentation using a variety of carbon sources. These carbon sources provide the necessary substrates for microorganisms to metabolize and convert into lactic acid. Lactic acid production from as a cheap carbon source have been reported by several authors²³⁻²⁵. Some of the common carbon sources for lactic acid production include:
- **3.1.Glucose**: Glucose is one of the most commonly used carbon sources for lactic acid fermentation²⁶⁻²⁷. It can be derived from various feedstocks, such as corn starch, sugarcane, or cellulosic biomass. Efficient production of l-lactic acid using co-feeding strategy based on cane molasses/ glucose carbon sources²⁸.
- **3.2.Lactose**: Lactose, a sugar found in milk, can also be used as a carbon source for lactic acid production have also been reported²⁹. Lactic acid bacteria, such as Lactobacillus, are often used for this purpose³⁰.
- **3.3.Molasses**: Recently much of the work have been carried out by using molasses, a byproduct of sugar refining processes, is rich in sugars and serves as an economical



carbon source for lactic acid fermentation. Cane molasses, a waste from sugar manufacturing processes, is hopeful to be utilized as cheap carbon source for L-lactic acid fermentation ³¹⁻³³.

- **3.4.Starch**: Starch from sources like corn, cassava, or potatoes can be enzymatically hydrolyzed into glucose or maltose, which can be used as a carbon source for the production of lactic acid³⁴⁻³⁶.
- **3.5.Agro-industrial Residues**: Various agricultural and food processing residues, such as wheat bran, rice bran, and fruit pomace, can be used as carbon sources for lactic acid production after appropriate pretreatment and enzymatic hydrolysis. Relatively to substrate sources, worldwide there is a lot of interesting agro-industrial waste or sub-products with a lower value, which can be fermented by several organisms ³⁷⁻³⁹.
- **3.6.Cellulosic Biomass**: Cellulosic materials, like agricultural residues and dedicated energy crops, can be broken down into fermentable sugars, including glucose and xylose, and used for lactic acid production⁴⁰⁻⁴³.
- **3.7.Glycerol**: Glycerol, a byproduct of biodiesel production, can serve as a carbon source for lactic acid fermentation, especially by certain strains of lactic acid bacteria⁴⁴⁻⁴⁵.
- **3.8.Whey**: Whey, a byproduct of cheese and yogurt production, contains lactose and can be used as a carbon source for lactic acid production⁴⁶⁻⁴⁹.
- **3.9.Lignocellulosic Biomass**: Advanced processes involving enzymatic hydrolysis can break down lignocellulosic biomass, such as wood or agricultural residues, into fermentable sugars like glucose and xylose for lactic acid production⁵⁰⁻⁵⁶.

The choice of carbon source depends on factors such as cost, availability, and the specific microorganism or fermentation process being used. Different strains of lactic acid



IJFANS INTERNATIONAL JOURNAL OF FOOD AND NUTRITIONAL SCIENCES ISSN PRINT 2319 1775 Online 2320 7876

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bacteria or other microorganisms may have varying preferences for carbon sources, and the selection of the most suitable source can significantly impact the efficiency and economics of lactic acid production.

Carbon Source	Description
Glucose	Commonly used from various feedstocks.
Lactose	Found in milk, utilized by lactic acid bacteria.
Molasses	Byproduct of sugar refining, economical source.
Starch	Hydrolyzed into glucose or maltose for use.
Agro-Industrial Residues	Residues like wheat bran, rice bran, etc.
Cellulosic Biomass	Agricultural and dedicated energy crops.
Glycerol	Byproduct of biodiesel production.
Whey	Byproduct of cheese and yogurt production.
Lignocellulosic Biomass	Requires enzymatic hydrolysis for sugars.

Molasses are by-products from sugar manufacturing, generally used as animal feed and for the production of bio-ethanol and yeasts⁵⁷. Sucrose is the most abundant sugar in their composition, but due to its high concentration, the viscosity of the liquid is important, resulting in an increase of the operating costs⁵⁸. The most common strain to ferment molasses is *Lactobacillus delbruecki*⁵⁹. Lactic acid production from cane molasses by *Lactobacillus delbruecki* NCIM 2025 in submerged condition: optimization of medium component by Taguchi DOE Methodology⁶⁰. Diptendu Sarkar et al⁶¹. and Arun Kumar etal⁶² explored the optimization of lactic acid production by adjusting various process parameters, such as medium pH, temperature, inoculum size, incubation time, and shaking speed⁶⁰.

Some of the work on the production of lactic acid from molasses has been reported and summarized in Table 2.



Organism	Lactic acid (g/L)	Yield g/g	Productivi ty g/(L/h)	Ref.
Lb. delbruckii NCIMB 8130	90	0.97	3.8	63
Lb. delbrueckii	88	-	-	63
Lb. delbrueckii subsp. delbrueckii Mutant Uc-3	166	-	4.15	64
Lb. delbrueckii	107	0.9	1.48	65
B. coagulans	168.3	0.88	2.1	66
E. faecalis RKY1	95.7	-	4.0	67
Lb.paracasei	169.9	-	1.42	68
E. coli	75	0.85	1.18	69
Microbial consortium CEE-DL15 Clostridium sensustricto (57.29%), Escherichia (34.22%), andEnterococcus (5.32%)	112.3	0.81	4.49	70
Bacillus coagulans	168.3	(0.88	2.1	71
Lb. delbrueckii sp. bulgaricus AU	20	0.45	-	72
Lb. delbrueckii sp. delbrueckii ATCC 9649	26	0.58	-	72
Lb. rhamnosus ATCC 7469	18	0.40	-	72

Table 2. Microbial production of lactic acid from molasses

4. Conclusion:

The review of the physical-chemical properties and microbial production of lactic acid provides valuable insights into the versatile and important compound, lactic acid. This organic acid has a wide range of applications in various industries, including food, pharmaceuticals, and bioplastics. Microbial production of lactic acid, often referred to as fermentation, is a sustainable and economically viable method. Lactic acid bacteria, such as Lactobacillus and Lactococcus species, are commonly used for this purpose.

The choice of substrates for microbial production is diverse. These substrates can include carbohydrates derived from various sources, including agricultural crops, lignocellulosic biomass, and agro-industrial residues. The selection of feedstock affects the sustainability and cost-effectiveness of lactic acid production. To improve lactic acid yield, researchers



employ various strategies such as strain improvement, process optimization, and genetic engineering. These approaches aim to enhance production efficiency and reduce byproducts. This review underscores the significance of lactic acid and its production processes in the context of green and sustainable chemistry.

5. References

- Abd Alsaheb, R. A., Aladdin, A., Othman, N. Z., Abd Malek, R., Leng, O. M., Aziz, R., & El Enshasy, H. A. (2015). Lactic acid applications in pharmaceutical and cosmeceutical industries. Journal of Chemical and Pharmaceutical Research, 7(10), 729-735.
- 2. Babilas, D., & Dydo, P. (2018). Selective zinc recovery from electroplating wastewaters by electrodialysis enhanced with complex formation. Separation and Purification Technology, 192, 419-428.
- EFSA Panel on Food Contact Materials, Enzymes and Processing Aids (CEP), Silano, V., Barat Baviera, J. M., Bolognesi, C., Brüschweiler, B. J., Chesson, A., Cocconcelli, P. S., Crebelli, R., Gott, D. M., Grob, K., & Lampi, E. (2018). Evaluation of the safety and efficacy of the organic acids lactic and acetic acids to reduce microbiological surface contamination on pork carcasses and pork cuts. Efsa Journal, 16(12), e05482.
- 4. Qi, X., Ren, Y., & Wang, X. (2017). New advances in the biodegradation of Poly (lactic) acid. International Biodeterioration & Biodegradation, 117, 215-223.
- Mohania, D., Nagpal, R., Kumar, M., Bhardwaj, A., Yadav, M., Jain, S., Marotta, F., Singh, V., Parkash, O. M., & Yadav, H. (2008). Molecular approaches for identification and characterization of lactic acid bacteria. Journal of Digestive Diseases, 9(4), 190-198.
- Eş, I., Khaneghah, A. M., Barba, F. J., Saraiva, J. A., Sant'Ana, A. S., & Hashemi, S. M. (2018). Recent advancements in lactic acid production—a review. Food Research International, 107, 763-770.
- 7. Tian, X., Chen, H., Liu, H., & Chen, J. (2021). Recent advances in lactic acid production by lactic acid bacteria. Applied Biochemistry and Biotechnology, 1-21.
- 8. Li, C., Gao, M., Zhu, W., Wang, N., Ma, X., Wu, C., & Wang, Q. (2021). Recent advances in the separation and purification of lactic acid from fermentation broth. Process Biochemistry, 104, 142-151.
- Ahmad, A., Banat, F., & Taher, H. (2020). A review on the lactic acid fermentation from low-cost renewable materials: Recent developments and challenges. Environmental Technology & Innovation, 20, 101138.



- Li, Y., Bhagwat, S. S., Cortés-Peña, Y. R., Ki, D., Rao, C. V., Jin, Y. S., & Guest, J. S. (2021). Sustainable lactic acid production from lignocellulosic biomass. ACS Sustainable Chemistry & Engineering, 9(3), 1341-1351.
- Esquivel-Hernández, D. A., García-Pérez, J. S., López-Pacheco, I. Y., Iqbal, H. M., & Parra-Saldívar, R. (2022). Resource recovery of lignocellulosic biomass waste into lactic acid-Trends to sustain cleaner production. Journal of Environmental Management, 301, 113925.
- 12. Lin, H. T., Huang, M. Y., Kao, T. Y., Lu, W. J., Lin, H. J., & Pan, C. L. (2020). Production of lactic acid from seaweed hydrolysates via lactic acid bacteria fermentation. Fermentation, 6(1), 37.
- Nwamba, M. C., Sun, F., Mukasekuru, M. R., Song, G., Harindintwali, J. D., Boyi, S. A., & Sun, H. (2021). Trends and hassles in the microbial production of lactic acid from lignocellulosic biomass. Environmental Technology & Innovation, 21, 101337.
- Karnaouri, A., Asimakopoulou, G., Kalogiannis, K. G., Lappas, A., & Topakas, E. (2020). Efficient d-lactic acid production by Lactobacillus delbrueckii subsp. bulgaricus through conversion of organosolv pretreated lignocellulosic biomass. Biomass and Bioenergy, 140, 105672.
- 15. van Lieshout, G. P. (1992). The Physical Properties of Lactic Acid and Derivatives: A literature review.
- 16. Mohanty, J. N., Das, P. K., Nanda, S., Nayak, P., & Pradhan, P. (2015). Comparative analysis of crude and pure lactic acid produced by Lactobacillus fermentum and its inhibitory effects on spoilage bacteria. The Pharma Innovation Journal, 3(11), 38-42.
- Vaidya, A. N., Pandey, R. A., Mudliar, S., Kumar, M. S., Chakrabarti, T., & Devotta, S. (2005). Production and recovery of lactic acid for polylactide—an overview. Critical Reviews in Environmental Science and Technology, 35(5), 429-467.
- Castillo Martinez, F. A., Balciunas, E. M., Salgado, J. M., Domínguez González, J. M., Converti, A., & Oliveira, R. P. de S. (2013). Lactic acid properties, applications and production: A review. Trends in Food Science & Technology, 30(1), 70–83.
- 19. Komesu, A., de Oliveira, J. A., da Silva Martins, L. H., Maciel, M. R., & Maciel Filho, R. (2017). Lactic acid production to purification: a review. BioResources, 12(2), 4364-4383.
- Pillin, I., Montrelay, N., Bourmaud, A., & Grohens, Y. (2008). Effect of thermomechanical cycles on the physico-chemical properties of poly (lactic acid). Polymer Degradation and Stability, 93(2), 321-328.
- 21. Ploypetchara, N., Suppakul, P., Atong, D., & Pechyen, C. (2014). Blend of polypropylene/poly (lactic acid) for medical packaging application: physicochemical, thermal, mechanical, and barrier properties. Energy Procedia, 56, 201-210.



- 22. Emel'Yanenko, V. N., Verevkin, S. P., Schick, C., Stepurko, E. N., Roganov, G. N., & Georgieva, M. K. (2010). The thermodynamic properties of S-lactic acid. Russian Journal of Physical Chemistry A, 84, 1491-1497.
- Petrut, S., Rusu, E., Tudorache, I. S., Pelinescu, D., Sarbu, I., Stoica, I., & Vassu, T. (2019). Influence of various carbon sources on growth and biomass accumulation of some lactic acid bacteria strains. Pharmaceuticals, 4, 5.
- 24. Juturu, V., & Wu, J. C. (2016). Microbial production of lactic acid: the latest development. Critical Reviews in Biotechnology, 36(6), 967-977.
- 25. Xu, K., & Xu, P. (2014). Efficient production of L-lactic acid using co-feeding strategy based on cane molasses/glucose carbon sources. Bioresource Technology, 153, 23-29.
- Petrut, S., Rusu, E., Tudorache, I. S., Pelinescu, D., Sarbu, I., Stoica, I., & Vassu, T. (2019). Influence of various carbon sources on growth and biomass accumulation of some lactic acid bacteria strains. Pharmaceuticals, 4, 5.
- Abdel-Rahman, M. A., Tashiro, Y., & Sonomoto, K. (2011). Lactic acid production from lignocellulose-derived sugars using lactic acid bacteria: overview and limits. Journal of Biotechnology, 156(4), 286-301.
- Reddy, G., Altaf, M. D., Naveena, B. J., Venkateshwar, M., & Kumar, E. V. (2008). Amylolytic bacterial lactic acid fermentation—a review. Biotechnology Advances, 26(1), 22-34.
- 29. Xu, K., & Xu, P. (2014). Efficient production of L-lactic acid using co-feeding strategy based on cane molasses/glucose carbon sources. Bioresource Technology, 153, 23-29.
- Petrut, S., Rusu, E., Tudorache, I. S., Pelinescu, D., Sarbu, I., Stoica, I., & Vassu, T. (2019). Influence of various carbon sources on growth and biomass accumulation of some lactic acid bacteria strains. Pharmaceuticals, 4, 5.
- Ahmad, A., Banat, F., & Taher, H. (2020). A review on the lactic acid fermentation from low-cost renewable materials: Recent developments and challenges. Environmental Technology & Innovation, 20, 101138.
- 32. Njokweni, S. G., Steyn, A., Botes, M., Viljoen-Bloom, M., & van Zyl, W. H. (2021). Potential valorization of organic waste streams to valuable organic acids through microbial conversion: a South African case study. Catalysts, 11(8), 964.
- 33. Verma, D. K., Patel, A. R., Thakur, M., Singh, S., Tripathy, S., Srivastav, P. P., Chavez-Gonzalez, M. L., Gupta, A. K., & Aguilar, C. N. (2021). A review of the composition and toxicology of fructans, and their applications in foods and health. Journal of Food Composition and Analysis, 99, 103884.
- 34. Jagatee, S., Behera, S., Dash, P. K., Sahoo, S., & Mohanty, R. C. (2015). Bioprospecting starchy feedstocks for bioethanol production: a future perspective. JMRR, 3, 24-42.



- 35. Reddy, G., Altaf, M. D., Naveena, B. J., Venkateshwar, M., & Kumar, E. V. (2008). Amylolytic bacterial lactic acid fermentation—a review. Biotechnology Advances, 26(1), 22-34.
- 36. Sharma, A., Singh, S., Khare, S. K., Sharma, A., Tiwari, R., Nain, L. (2022). Green lactic acid production using low-cost renewable sources and potential applications. In Production of Top 12 Biochemicals Selected by USDOE from Renewable Resources (pp. 345-365). Elsevier.
- 37. Panesar, P. S., & Kaur, S. (2015). Bioutilization of agro- industrial waste for lactic acid production. International Journal of Food Science & Technology, 50(10), 2143-2151.
- Alexandri, M., Schneider, R., Mehlmann, K., Venus, J. (2019). Recent advances in dlactic acid production from renewable resources: Case studies on agro-industrial waste streams. Food Technology and Biotechnology, 57(3), 293-304.
- Mladenović, D., Pejin, J., Kocić-Tanackov, S., Radovanović, Ž., Djukić-Vuković, A., Mojović, L. (2018). Lactic acid production on molasses enriched potato stillage by Lactobacillus paracasei immobilized onto agro-industrial waste supports. Industrial Crops and Products, 124, 142-148.
- 40. Kumar, M. N., Gialleli, A. I., Masson, J. B., Kandylis, P., Bekatorou, A., Koutinas, A. A., Kanellaki, M. (2014). Lactic acid fermentation by cells immobilized on various porous cellulosic materials and their alginate/poly-lactic acid composites. Bioresource Technology, 165, 332-335.
- 41. Shen, X., & Xia, L. (2006). Lactic acid production from cellulosic material by synergetic hydrolysis and fermentation. Applied Biochemistry and Biotechnology, 133, 251-262.
- 42. Adsul, M. G., Varma, A. J., & Gokhale, D. V. (2007). Lactic acid production from waste sugarcane bagasse derived cellulose. Green Chemistry, 9(1), 58-62.
- 43. Shumigin, D., Tarasova, E., Krumme, A., & Meier, P. (2011). Rheological and mechanical properties of poly (lactic) acid/cellulose and LDPE/cellulose composites. Materials Science, 17(1), 32-37.
- 44. Hong, A. A., Cheng, K. K., Peng, F., Zhou, S., Sun, Y., Liu, C. M., & Liu, D. H. (2009). Strain isolation and optimization of process parameters for bioconversion of glycerol to lactic acid. Journal of Chemical Technology & Biotechnology, 84(10), 1576-1581.
- 45. Jodłowski, G. S., & Strzelec, E. (2021). Use of glycerol waste in lactic acid bacteria metabolism for the production of lactic acid: State of the art in Poland. Open Chemistry, 19(1), 998-1008.
- Pescuma, M., de Valdez, G. F., & Mozzi, F. (2015). Whey-derived valuable products obtained by microbial fermentation. Applied Microbiology and Biotechnology, 99, 6183-6196.



- 47. Turner, T. L., Kim, E., Hwang, C., Zhang, G. C., Liu, J. J., & Jin, Y. S. (2017). Conversion of lactose and whey into lactic acid by engineered yeast. Journal of Dairy Science, 100(1), 124-128.
- 48. Sayed, W. F., Salem, W. M., Sayed, Z. A., & Abdalla, A. K. (2020). Production of lactic acid from whey permeates using lactic acid bacteria isolated from cheese. SVU-International Journal of Veterinary Sciences, 3(2), 78-95.
- 49. Zotta, T., Solieri, L., Iacumin, L., Picozzi, C., & Gullo, M. (2020). Valorization of cheese whey using microbial fermentations. Applied Microbiology and Biotechnology, 104(7), 2749-2764.
- 50. Taherzadeh, M. J., & Karimi, K. (2007). Enzymatic-based hydrolysis processes for ethanol from lignocellulosic materials: A review. BioResources, 2(4), 707-738.
- 51. Vasić, K., Knez, Ž., & Leitgeb, M. (2021). Bioethanol production by enzymatic hydrolysis from different lignocellulosic sources. Molecules, 26(3), 753.
- 52. Tandon, G. (2015). Bioproducts from residual lignocellulosic biomass. In Advances in Biotechnology (pp. 52-75).
- Swamba, M. C., Sun, F., Mukasekuru, M. R., Song, G., Harindintwali, J. D., Boyi, S. A., & Sun, H. (2021). Trends and hassles in the microbial production of lactic acid from lignocellulosic biomass. Environmental Technology & Innovation, 21, 101337.
- 54. Yankov, D. (2022). Fermentative lactic acid production from lignocellulosic feedstocks: from source to purified product. Frontiers in Chemistry, 10, 823005.
- 55. Vu, H. P., Nguyen, L. N., Vu, M. T., Johir, M. A., McLaughlan, R., & Nghiem, L. D. (2020). A comprehensive review on the framework to valorize lignocellulosic biomass as biorefinery feedstocks. Science of the Total Environment, 743, 140630.
- 56. Kumar, A. K., & Sharma, S. (2017). Recent updates on different methods of pretreatment of lignocellulosic feedstocks: a review. Bioresources and Bioprocessing, 4(1), 1-9.
- 57. Beigbeder, J. B., de Medeiros Dantas, J. M., & Lavoie, J. M. (2021). Optimization of yeast, sugar, and nutrient concentrations for high ethanol production rate using industrial sugar beet molasses and response surface methodology. Fermentation, 7(2), 86.
- 58. Doldolova, K., Bener, M., Lalikoğlu, M., Aşçı, Y. S., Arat, R., & Apak, R. (2021). Optimization and modeling of microwave-assisted extraction of curcumin and antioxidant compounds from turmeric by using natural deep eutectic solvents. Food Chemistry, 353, 129337.
- 59. Bath, S. M., & Srivastava, S. K. (2008). Lactic acid production from cane molasses by Lactobacillus delbruecki NCIM 2025 in submerged condition: optimization of medium component by Taguchi DOE Methodology. Food Biotechnology, 22, 115-139.



- 60. Bhatt, S. M., & Srivastava, S. K. (2008). Lactic acid production from cane molasses by Lactobacillus delbrueckii NCIM 2025 in submerged condition: optimization of medium component by Taguchi DOE methodology. Food Biotechnology, 22(2), 115-139.
- 61. Sarkar, D., & Paul, G. (2019). A STUDY ON OPTIMIZATION OF LACTIC ACID PRODUCTION FROM WHEY BY LACTOBACILLUS SP ISOLATED FROM CURD SAMPLE. Research Journal of Life Sciences, Bioinformatics, Pharmaceutical and Chemical Sciences, 5(2), 816-824.
- 62. Alam, M. T., Mahato, A. K., Kumari, L., & Singh, R. S. (2021). Fermentative study on optimization of lactic acid production from cane sugar by Lactobacillus spp. European Journal of Molecular & Clinical Medicine, 8(2), 712-723.
- 63. Kotzamanidis, C., Roukas, T., & Skaracis, G. (2002). Optimization of lactic acid production from beet molasses by Lactobacillus delbrueckii NCIMB 8130. World Journal of Microbiology and Biotechnology, 18, 441-444.
- 64. Dumbrepatil, A., Adsul, M., Chaudhari, S., Khire, J., & Gokhale, D. (2008). Utilization of molasses sugar for lactic acid production by Lactobacillus delbrueckii subsp. delbrueckii mutant Uc-3 in batch fermentation. Applied and Environmental Microbiology, 74, 333-335.
- Abdel-Rahman, M. A., Tashiro, Y., & Sonomoto, K. (2013). Recent advances in lactic acid production by microbial fermentation processes. Biotechnology Advances, 31, 877-902.
- 66. Xu, K., & Xu, P. (2014). Efficient production of L-lactic acid using co-feeding strategy based on cane molasses/glucose carbon sources. Bioresource Technology, 153, 23-29.
- 67. Yun, J.-S., Wee, Y.-J., Kim, J.-N., & Ryu, H.-W. (2004). Fermentative production of dllactic acid from amylase-treated rice and wheat brans hydrolyzate by a novel lactic acid bacterium, Lactobacillus sp. Biotechnology Letters, 26, 1613-1616.
- Mladenović, D., Pejin, J., Kocić-Tanackov, S., Radovanović, Ž., Djukić-Vuković, A., & Mojović, L. (2018). Lactic acid production on molasses enriched potato stillage by Lactobacillus paracasei immobilized onto agro-industrial waste supports. Industrial Crops and Products, 124, 142-148.
- 69. Wang, Y. Z., Li, K. P., Huang, F., Wang, J. H., Zhao, J. F., Zhao, X., & Zhou, S. D. (2013). Engineering and adaptive evolution of Escherichia coli W for L-lactic acid fermentation from molasses and corn steep liquor without additional nutrients. Bioresource Technology, 148, 394-400.
- 70. Sun, Y., Xu, Z., Zheng, Y., Zhou, J., & Xiu, Z. (2019). Efficient production of lactic acid from sugarcane molasses by a newly microbial consortium CEE-DL15. Process Biochemistry, 81, 132-138.



- 71. Xu, K., & Xu, P. (2014). Efficient production of L-lactic acid using co-feeding strategy based on cane molasses/glucose carbon sources. Bioresource Technology, 153, 23-29. doi:10.1016/j.biortech.2013.11.057
- 72. Tiwari, K. P., Pandey, A., & Mishra, N. (1979). Lactic acid production from molasses by mixed population of Lactobacilli. Zentralbl Bakteriol Parasitenkd Infektionskr Hyg Zweite Naturwiss Abt Mikrobiol Landwirtsch Technol Umweltschutzes, 134, 544-546.

