

***In-vitro* Evaluation of Antibacterial Potential of Biocontrol Agents Against Pathogens of *Punica granatum* L.**

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ABSTRACT

Pomegranate (*Punica granatum* L.) is one of the highly valued table fruits, grown in various parts of the world as it has great commercial, medicinal and nutritive value. India is one of the major producers of pomegranates that are exported to various parts of the world and Maharashtra leads in pomegranate production in the country. Thus, in India it is a serious economic threat to pomegranate growers due to lack of effective control measures. Biological control is the most viable approach for the treatment of plant diseases. Thus, there is a need for a safe biocontrol agent. In view of this background, several samples of leaves, fruits, buds and fruits were collected from Pomegranate orchards of Maharashtra for the evaluation of bacterial population and isolation of *Trichoderma* species. Thus, to evaluate the possibility of their extendable use as biocontrol agents in pomegranate. Pomegranate crop is attacked by several bacterial and fungal pathogens. Thus, these diseases of pomegranate decrease the market value of fruits. Use of bactericides and fungicides is a conventional method to control infections which has hazardous impact on public health and these agrochemicals are not cost effective. Biological control is the most viable approach for the treatment of plant diseases. Thus, there is a need for a safe biocontrol agent and the *Trichoderma* fungi belong to this category. The biocontrol potential of different *Trichoderma* species Viz. *Trichoderma viride* VMB21, *Trichoderma reesei* VMB23 and *Trichoderma stercorarium* VMB25. has been demonstrated on Pomegranate bacterial pathogens Viz. *Xanthomonas axonopodis* VMB13, *Xanthomonas campestris* VMB15 and *Xanthomonas vesicatoria* VMB17.

Keywords: Pomegranate, *Xanthomonas* species, *Trichoderma* species, Plants extract, Antibacterial activity.

INTRODUCTION

Plant diseases need to be controlled to maintain the quality and abundance of food, feed, and fiber produced by growers around the world. Different approaches may be used to prevent, mitigate or control plant diseases. Beyond good agronomic and horticultural practices, growers often rely heavily on chemical fertilizers and pesticides. Such inputs to agriculture have contributed significantly to the spectacular

improvements in crop productivity and quality over the past 100 years. However, the environmental pollution caused by excessive use and misuse of agrochemicals, as well as fear-mongering by some opponents of pesticides, has led to considerable changes in people's attitudes towards the use of pesticides in agriculture. Today, there are strict regulations on chemical pesticide use, and there is political pressure to remove the most hazardous chemicals from the market. Additionally, the spread of plant diseases in natural ecosystems may prevent successful application of chemicals, because of the scale to which such applications might have to be applied. Consequently, some pest management researchers have focused their efforts on developing alternative inputs to synthetic chemicals for controlling pests and diseases. Among these alternatives are those referred to as biological control.

Biological control or biocontrol is defined as the use of organisms in the management of plant diseases. These organisms are referred to as biocontrol agents. In plant pathology, the term applies to the use of antagonistic microbes to suppress the diseases of plants. The use of host specific pathogens for the control of weeds also comes under this field. The biocontrol agents should be safe from humans and environment point of view.

The biocontrol potential of different *Trichoderma species* Viz. *Trichoderma viride* VMB21, *Trichoderma reesei* VMB23 and *Trichoderma stercorarium* VMB25. has been demonstrated on Pomegranate bacterial pathogens Viz. *Xanthomonas axonopodis* VMB13, *Xanthomonas campestris* VMB15 and *Xanthomonas vesicatoria* VMB17 by agar well diffusion assay.

MATERIALS AND METHODS

Preliminary identification and isolation of the pathogen from affected parts of Pomegranate plant

Ooze test was performed as per description of Sharma *et al.*, (2010) which is based on the microscopic observations for confirmation of presence of the pathogen in infected leaves. Pomegranate leaves which show typical symptoms of oily spot (bacterial blight) were collected from the field. In this test, the infected leaves were washed with the sterile distilled water. Further, all the selected infected leaves were sterilized with 0.1% mercuric chloride (HgCl₂) for 10 minutes and followed by repeatedly washing with sterile distilled water and next blot dried. Then a fresh young lesion containing a leaf was selected. The small bits of infected tissue were cut down from affected portions of plants such as leaves and pericarp of the fruits with the help of a sharp sterile scalpel. Then it was placed in a drop of sterile saline taken on a glass slide and observed from cut ends under the high power objective of the compound microscope. Jet of bacterial cells started oozing out from the cut pieces of the tissue in water were observed under a microscope called 'ooze' (Schaad, 1992).

Further, suspension of bacteria prepared from infected tissues taken from affected leaves, twigs and fruits were used for isolation of well separated bacterial colonies on suitable medium. Singh *et al.*, (2015) stated the disease diagnosis depends on visual symptoms on plant parts and ooze test.

Isolation of bacteria exudes out from ooze

The leaf having the infected part was washed and further crushed in sterile distilled water. The extract obtained was then streaked inoculated on the surface of a sterile Glucose Yeast Extract Calcium Carbonate Agar (GYECC Agar) (Yenjerappa, 2009). The plate was further incubated at 30⁰C for 3 days. As per the literature, a well isolated, gummy, mucous, yellow colored colony was selected for further study (Yenjerappa, 2009).

Isolation of *Trichoderma species*

Soil samples were collected from different ecological habitats of Latur district, (India) for the isolation of *Trichoderma spp.* These soil samples were brought to the laboratory and stored in a refrigerator at 4⁰C until used. Four-fold serial dilutions of each soil sample were prepared in sterilized distilled water. 0.5 ml of this diluted sample was poured on the surface of Potato Dextrose Agar (Elad *et al.*, 1982). Plates were incubated at 28 ± 2⁰C for 96 hours. Morphologically distinct colonies appearing on the agar plates were purified on Potato Dextrose Agar (PDA). The purified promising isolates were further preserved at 4⁰C and used during the course of study.

Preparation of Solvent Extracts of Selected Medicinal Plants

Twenty grams of dried powder of each plant was filled in the thimble and extracted consecutively with the solvents *Viz.* methanol, ethanol and chloroform by using a Soxhlet extractor (Soxhlet Complete Borosil, Code 3840) for 48 hours. All these extracts were separately prepared in Soxhlet extractor. Further, all these extracts were concentrated using a rotary flash evaporator (Superfit Rotary Vacuum Flash Unit PBU-6D) and preserved at 4⁰C in airtight bottles for further use. Afterwards, all these extracts were subjected to antibacterial activity assay (Raghavendra *et al.*, 2006).

Determination of Antibacterial Activity

The antibacterial activity of aqueous extracts, solvent extracts and the mixture of all plant extracts under study were determined by agar well diffusion method (Cruickshank *et al.*, 1975) on the sterile nutrient glucose agar medium. Cell density was adjusted to 10⁶ -10⁷ CFU/ml on the basis when culture reaches 0.1 optical units at 600nm with spectrophotometer (Schaad, 1992). The inoculum containing 10⁶-10⁷ CFU / ml of 72 hours old culture of *Xanthomonas axonopodis* pv. *Punicae* VMB13, *Xanthomonas campestris* VMB15 and *Xanthomonas vesicatoria* VMB17 was spread inoculated on medium with the help of sterile glass spreader. Wells were prepared in sterile nutrient glucose agar plate by using sterile cork borer (5mm). 100µl of aqueous extract and solvent extract was put in the wells prepared in the inoculated plates

RESULTS AND DISCUSSIONS

Trichoderma is a soil-borne filamentous fungus that is most widely used as a biocontrol agent; since it is capable of parasitising several plant pathogenic fungi. *T. harzianum*, *T. viride* and *T. asperellum* have been mostly reported as biocontrol agents against plant pathogens (Harman *et al.*, 2004, 2011; Kumar *et al.*, 2012; Cuervo-Parra *et al.*, 2011).

Table 1: *In Vitro* Evaluation of Antibacterial Potential of Biocontrol Agents against Bacterial Pathogens of Pomegranate Diseases

| Bacterial Pathogens | <i>Trichoderma viride</i> VMB21 | <i>Trichoderma reesei</i> VMB23 | <i>Trichoderma</i> <i>stercorarium</i> VMB25 |
|--------------------------------------|------------------------------------|------------------------------------|---|
| | Zone of Growth Inhibition (mm) | | |
| <i>Xanthomonas axonopodis</i> VMB13 | 45 | 40 | 39 |
| <i>Xanthomonas campestris</i> VMB15 | 42 | 35 | 40 |
| <i>Xanthomonas vesicatoria</i> VMB17 | 41 | 38 | 31 |

***In Vitro* Evaluation of Biocontrol Agents against Bacterial Pathogens**

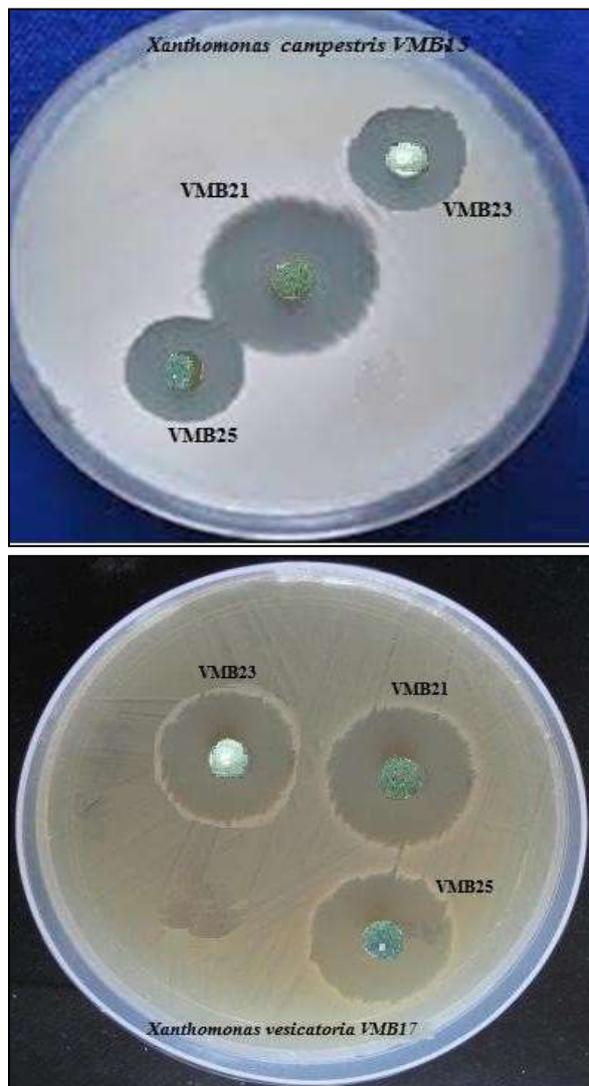
Trichoderma species are frequent inhabitants of the rhizosphere and contribute to control of many soil borne plant diseases which were caused by fungi. The potential of different *Trichoderma* species used as biological agents of plant diseases have been known since the 1930s. Therefore, three species of *Trichoderma* were selected for their antifungal activity against the targeted pathogens as follows.

- 1) *Trichoderma viride* VMB21
- 2) *Trichoderma reesei* VMB23
- 3) *Trichoderma stercorarium* VMB25.

Three Biocontrol agents *Viz.* *Trichoderma viride* VMB21, *Trichoderma reesei* VMB23 and *Trichoderma stercorarium* VMB25 were tested against three bacterial pathogens *Viz.* *Xanthomonas axonopodis* VMB13, *Xanthomonas campestris* VMB15, *Xanthomonas vesicatoria* VMB17 which were isolated from diseased Pomegranate leaves, stem and fruits. Zones of growth inhibition were measured and calculated. The results were tabulated in **Table 1**.

Among the three bioagents tested (**Table 1**), *Trichoderma viride* VMB21 showed the maximum inhibition of the test pathogens.



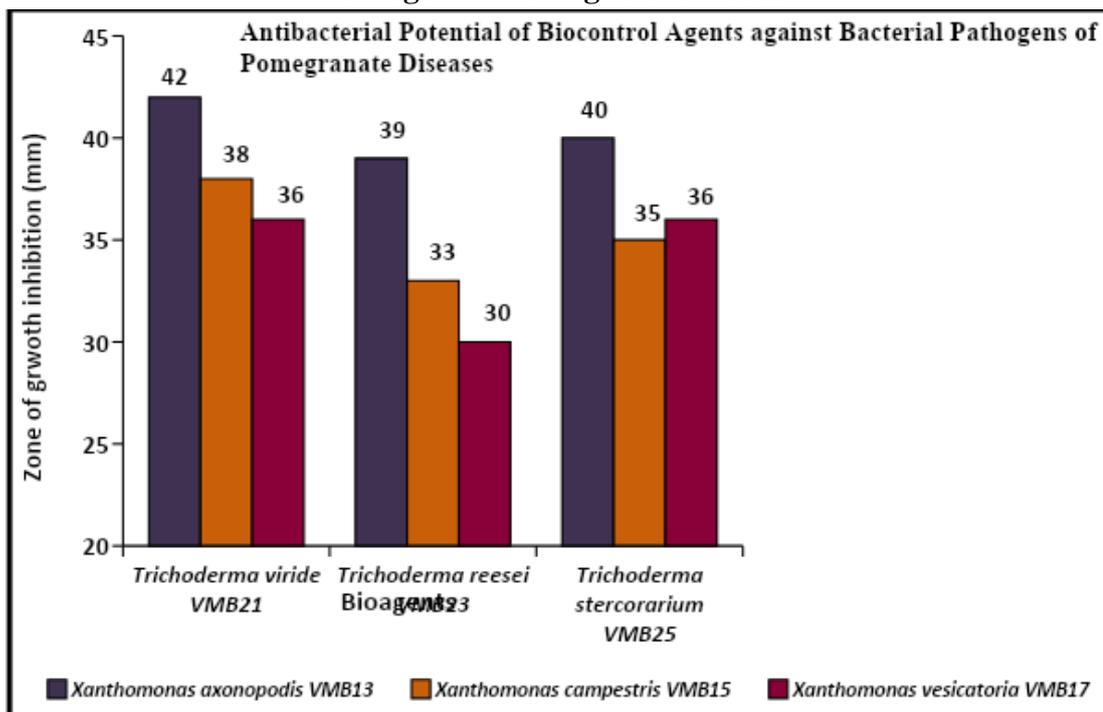


Photoplate:1 Antibacterial Potential of Biocontrol Agents against Bacterial Pathogens of Pomegranate Disease

Evaluation of Efficacy of Biocontrol agents against Bacterial Pathogen

In the light of present day constraints with the utilization of chemical pesticides in the plant disease management, the biological control as an alternate option is gaining very much importance and awareness since the approach is eco-friendly and cost effective. Under biological control of plant diseases, different antagonistic organisms have been identified, which fight against the pathogens by a range of mechanisms *Viz.*, competition, antibiosis, siderophore production, lysis, etc. In the present investigation *Trichoderma viride* showed maximum inhibition of the three bacterial pathogens *Viz.* *Xanthomonas axonopodis* VMB13, *Xanthomonas campestris* VMB15 and *Xanthomonas vesicatoria* VMB17.

Figure 1: Antibacterial Potential of Biocontrol Agents against Bacterial Pathogens of Pomegranate Disease



Antibacterial potential of three biocontrol agents viz. *Trichoderma viride* VMB21, *Trichoderma reesei* VMB23 and *Trichoderma stercorarium* VB25 against bacterial pathogens viz. *Xanthomonas axonopodis* VMB13, *Xanthomonas campestris* VMB15 and *Xanthomonas vesicatoria* VMB17 of pomegranate disease were evaluated. The results are depicted in **Figure 1**.

It is found from **Figure 1** that the three biocontrol agents viz. *Trichoderma viride* VMB21, *Trichoderma reesei* VMB23 and *Trichoderma stercorarium* VB25 shows the maximum zone of inhibition i.e. 42mm, 39mm and 40mm respectively to *Xanthomonas axonopodis* VMB13. The maximum zone of inhibition shown by the three bioagents *Trichoderma viride* VMB21, *Trichoderma reesei* VMB23 and *Trichoderma stercorarium* VMB25 to the *Xanthomonas campestris* VMB15 is 38mm, 33mm and 35mm respectively. Similarly, the zone of growth inhibition shown by the same bioagents to the *Xanthomonas vesicatoria* VMB17 is 36mm, 30mm and 36mm respectively.

Our results are in strong agreement with the results reported by Nurbailis *et al.*, (2019) in which they reported that *Trichoderma viride*, *Trichoderma harzianum* and *Trichoderma PP3* reducing the growth of *Xanthomonas axonopodis* pv. caused leaf blight disease on red onion.

Abhishek Gupta *et al.*, (2018) reported that the secondary metabolites of *Trichoderma viride* such as cellulase enzyme (this enzyme try to use as a bio-control agent) give strong antibacterial activity against plants pathogenic bacteria *Xanthomonas citri* which are cause disease in lemon plants.

Our results are in contrast with the results reported by Bhure *et al.*, (2019) in which they tested the antagonistic activity of three bioagents and confined that

Trichoderma harzianum gives maximum inhibition zone (22.86mm) followed by *Pseudomonas fluorescens* (17.20mm) and *Bacillus subtilis* (15.00mm).

Molecular Identification of Biocontrol Agents

Molecular identification of the biocontrol agents was carried out by 18s rRNA sequencing. Full length 18s rRNA region was amplified using the universal primers. The amplified PCR generated sequences were then analyzed at National Centre for Biotechnology information (NCBI), Bethesda, MD. www.ncbi.nlm.nih.gov/BLAST for closed homology using BLASTn algorithm.

The related sequences for fungal isolates were downloaded from the NCBI database and then were aligned by using CLUSTAL X2 and Dambe multiple sequence alignment tool. The Phylogenetic evolutionary relationship was then inferred using the Kimura-2- parameter and Neighbor Joining Method analysis.

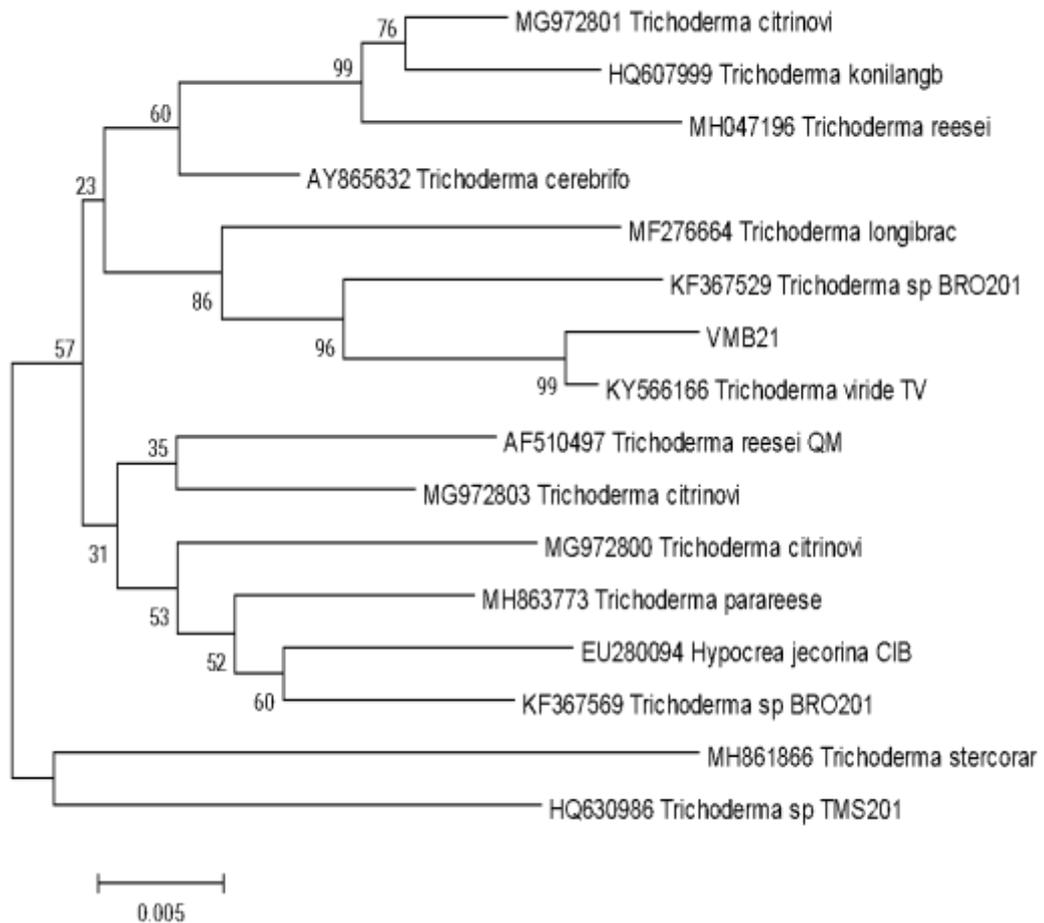


Figure 2: Phylogenetic placement of VMB21 (DDBJ Accession Number LC535970) based on 18S rRNA analysis

Phylogenetic tree of *Trichoderma viride VMB21*. Phylogenetic analysis of 18s rRNA gene sequence of *Trichoderma viride VMB21*. The percent numbers at the nodes indicate the levels of bootstrap support based on neighbor-joining analyses of 1,000 replicates. The scale bar (0.005) indicates the genetic distance.

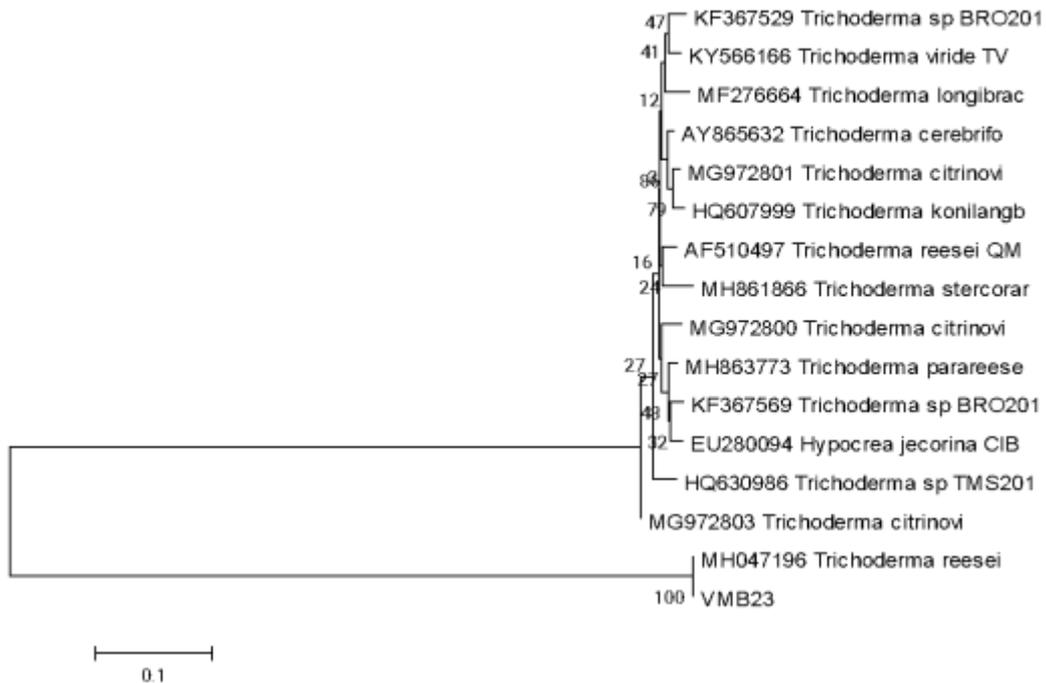


Figure 3: Phylogenetic placement of VMB23 (DDBJ Accession Number LC535971) based on 18S rRNA analysis

Phylogenetic tree of *Trichoderma reesei* VMB23. Phylogenetic analysis of 18S rRNA gene sequence of *Trichoderma reesei* VMB23. The percent numbers at the nodes indicate the levels of bootstrap support based on neighbor-joining analyses of 1,000 replicates. The scale bar (0.1) indicates the genetic distance.

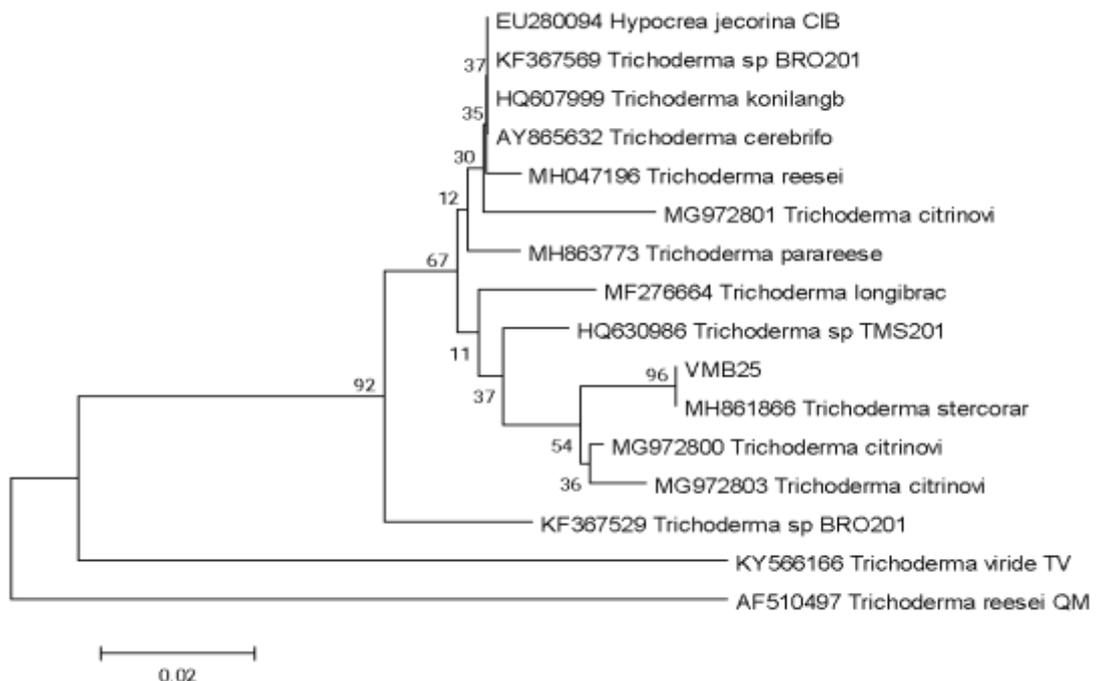


Figure 4: Phylogenetic placement of VMB25 (DDBJ Accession Number LC535972) based on 18S rRNA analysis

Phylogenic tree of *Trichoderma stercorearium* VMB25. Phylogenetic analysis of 18s rRNA gene sequence of *Trichoderma stercorearium* VMB25. The percent numbers at the nodes indicate the levels of bootstrap support based on neighbor-joining analyses of 1,000 replicates. The scale bar (0.02) indicates the genetic distance.

Shreeshail, S. *et al.*, (2015) used seven botanicals *Viz.* Neem leaf extract, Garlic bulb extract, Onion bulb extract, Datura leaf extract, Ocimum leaf extract, Eucalyptus leaf extract and Rhizome extract and four bioagents *Viz.* *Bacillus subtilis*, *Pseudomonas fluorescens*, *Trichoderma viride* and *Trichoderma harzianum* were evaluated *In Vitro* against the causal agent of wilt of pomegranate *i.e.* *Ceratocystis fimbriata*. The results of their study showed that among four bioagents *Trichoderma harzianum* and *Trichoderma viride* showed 100%inhibition of the test fungus *i.e.* *Ceratocystis fimbriata* followed by *Pseudomonas fluorescens* (42.33%) within 4 days and completely inhibited the perithecium production with growth over the pathogen.

Our results are in contrast with the results reported by Shreeshail, S. *et al.*, (2015) in that among the different bio agents *Trichoderma harzianum* showed the maximum inhibition of the test fungus *i. e.* the causal agent of wilt of pomegranate *Ceratocystis fimbriata* (100%) but remains on par with *Trichoderma viride* (100%).

Antibiotic-mediated suppression is the major mechanism that is noticed during biocontrol. Biocontrol agents produce one or several antimicrobial compounds that inhibit the growth of several plant pathogens. For example, *Bacillus cereus* strain is known to produce zwittermycin (Silo-Suh *et al.*, 1994) and kanosamine (Silo-Suh *et al.*, 1998). The ability to produce multiple antibiotics enables the biocontrol agents to inhibit a wide range of plant pathogens, which enhances the efficacy of biological control. *Pseudomonas putida* WCS358r, a genetically engineered strain produces phenazine and 2, 4-Diacetylphloroglucinol (DAPG), which exhibits improved suppression of plant diseases in fields (Glandorf *et al.*, 2001).

CONCLUSION

In all these interactions, the pathogens are antagonized by other organisms which indicate that there are several mechanisms involved in biological control. These mechanisms are hyperparasitism or predation, production of antibiotics, lytic enzymes, unregulated waste products and induction of host resistance. In hyperparasitism or predation, a specific biocontrol agent kills the pathogens by directly attacking it. Biocontrol agents produce one or several antimicrobial compounds that inhibit the growth of several plant pathogens. The ability to produce multiple antibiotics enables the biocontrol agents to inhibit a wide range of plant pathogens, which enhances the efficacy of biological control. The biocontrol potential of different *Trichoderma species Viz.* *Trichoderma viride* VMB21, *Trichoderma reesei* VMB23 and *Trichoderma stercorearium* VMB25 has been effectively demonstrated on Pomegranate bacterial pathogens *Viz.* *Xanthomonas axonopodis* VMB13, *Xanthomonas campestris* VMB15 and *Xanthomonas vesicatoria* VMB17.

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