

# Green Nanotechnology and Biological Synthesis of Gold Nanoparticles from Various Plant Extracts and Their Biological Applications

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## Abstract:

Nanotechnology is a trending area in research field for studying particles at nano scales and it is used over conventional methods and gives size dependent parcels of the functional accoutrements. The present review includes study on green technology for the synthesis of gold nanoparticles from various extracts of different plant parts. The green nanotechnology field that provides sustainability in various implementations. The nanomaterial synthesis using conventional nanotechnology involves the release of poisonous nanomaterials that creates the issue of nanotoxicity but the green nanotechnology helps in derogation of the risk and hazards due to the same [17]. The gold nanoparticles using plants extracts because of their diversity, sustainability and eco-friendly nature. It is concluded that gold nanoparticles synthesized using plants extract such as leaf, root, seed and fruit extracts have antioxidant and antimicrobial activity. Due to their SPR frequency, biocompatibility and tunable property gold nanoparticles has great application in cancer treatment, biosensor, drug delivery and enhancement of plant growth.

## 1. INTRODUCTION:-







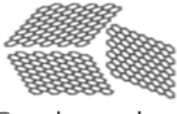

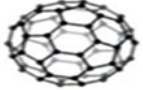
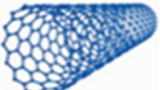
### 1.1. Nanotechnology and Nanoparticles:-

The word nanotechnology is taken from a greek word “nano” that means “dwarf” [1]. A Nanotechnology is an arising area of exploration which has an eventuality in relief of conventional micron technologies and gives size dependent parcels of the functional accoutrements. It confines to manufacturing at nano level, nanomaterials, and operation of physical, chemical and natural systems at scales from individual motes to submicron confines and also the fusion of the performing nanostructures into large structures [2]. Nanotechnology is concerned with systems and materials, the ingredients and structures of which due to their nanoscale size represent significantly better chemical, physical, and biological properties [3].

Nanotechnology basically deals with design, production and characterization of nanosized particles.

Nanomaterials are classified into different classes on the basis of dimension, as shown in **Table 1**. The different classes of nanomaterials, zero dimension, one dimension, two dimension, and three dimension. Zero dimension nanomaterials includes noble metal nanoparticles with size range of 1-100 nanometers such as Au Nps, AgNPs, Fe NPs, Cu NPs, etc., magnetic nanoparticles are composed of materials with high saturation magnetization such as Fe, Ni etc., fullerenes, and quantum dots such as graphene quantum dots, carbon quantum dots, inorganic quantum dots etc. [4].

**Table 1:- Classification of Gold Nanomaterials on the Basis of Dimension**

NANOMATERIALS			
0 Dimensional	1 Dimensional	2 Dimensional	3 Dimensional
 <u>Nanoparticles</u>	 <u>Nanowires</u>	 <u>Nanoplates</u>	 <u>Nanocrystals</u>
 <u>Quantum dots</u>	 <u>Nanorods</u>	 <u>Graphene sheets</u>	 <u>Liposome</u>
 <u>Fullerenes</u>	 <u>Nanotubes</u>		

One dimensional nanomaterial has two quantum-confined directions and one unconfined direction. One dimensional nanomaterials includes nanotubes, nanofibres, nanowires, and nanorods [5]. One dimensional nanomaterial have large length-to-diameter ratio, so they contain unique chemical, electrical, magnetic, and mechanical properties. For example nanowires such as zinc oxide nanowires, copper nanowires, silver nanowires etc. can be used for construction of wearable soft electronics [6]. Carbon nanorods, silicon nanotubes, silicon nanorods have been proposed as electrodes for energy storage [7].

Two dimensional (2D) nanomaterials have structure with uniformity generally ranging from atomic/molecular level to few hundreds of nanometers. Graphene is an example of soft 2D nanomaterial; it is single layered graphite that has close-packed conjugated hexagonal

carbon lattices.  $B_xC_yN_z$  is also an example of soft 2D nanomaterial. 2D  $B_xC_yN_z$  nanomaterials, includes hexagonal boron-carbon-nitrogen, hexagonal boron nitride and carbon nitride nanosheets etc. [8].

Multi-nanolayered bundles of nanowires or nanorods or ordered aggregation of nanoparticles give rise to 3D nanomaterials. Hierarchical h-BN has ultra thin 3dimensional foam structure [9]. Flower like nickel-cobalt oxide nanomaterials have also been synthesized [10]. Three dimensional carbon nanostructures has also been made by connecting two dimensional graphene and one dimensional carbon nanotubes. 3D G-CNT-Pd electrode material has an excellent application as supercapacitors [11].

The nanoparticle is principally a small particle ranging between 1 and 100 nanometers (nm) in diameter [12]. Surface area to volume ratio is unexpectedly high in NPs [13]. There is a prompt increase in the nanoparticles's use due to their immense application like in bio-imaging, cancer treatment, cosmetic surgery, biomedicine, DNA binding, solar cells, protein purification [14]. The problem of low solubility of herbal drugs are also solved by nanomedicine [15]. Herbal drugs with nanoparticles have more advantages and efficient than conventional forms of drugs [16]. In pharmacological studies, nanoparticles are also used as a carrier which provides specific targeting and drug delivery thus enhancing the bio-availability.

## 1.2. Green Nanotechnology:-

Green nanotechnology is related to the field that provides sustainability in various implementations. The main aim of it is the development of non threatening methods for the NPs synthesis. The nanomaterial synthesis involve the release of poisonous nanomaterials that creates the issue of nanotoxicity, the green nanotechnology helps in derogation of the risk and hazards due to the same [17].

It pursuit to the evaluate the synthetic styles that help to reduce the use of toxic chemicals with intensifying the capability of current synthesis techniques [18]. Green nanotechnology concern with the production of green products. Green products are those whose synthesis start from nontoxic precursor, use of natural source as raw material, involve less energy consumption, use of green solvents and no toxic disposal [19]. The aim of green nanotechnology is to develop environment friendly nanoparticles, develop green methods for large scale nanoparticle production, Discover efficient approaches for using nanoparticles in the development of novel nano-devices [20].

## 1.3. Why we use Green Nanotechnology:-

There is an increasing problem of environmental pollution in today's world due to humanistic activities. Something which is greener and cleaner is more preferable. Green nano-products can be directly applied to check damage from known adulterants and incorporation into environmental technologies can remediate unsafe waste sites, treat pollutants, plant pathogens and related toxins, sense and examiner environmental adulterants, and clean and desalinate weakened water. We prefer green nanotechnology because it provides safe and energy efficient products, reduction in waste and greenhouse gas emissions and involve the use of renewable materials [21].



**Figure 1. Principles of Green Chemistry on which Green Nanotechnology Based**

Green nanotechnology work on the 12 principles of green chemistry (shown in [Figure 1.](#)) such as to design new nanomaterials to achieve economic, social, health, and environmental benefits [22]. The principles of green chemistry impeccably apply to green nanotechnology and nano-manufacturing processes to produce safe, cleaner and more sustainable nanomaterials.

#### **1.4. Gold Nanoparticles:-**

Gold is the quintessential noble element. In its bulk shape, gold's uses in jewelry, coinage, and electronics are widely recognized. Gold nanoparticles (Au NPs) have attracted extensive attention in the last decades due to their unique catalytic, photo physical and electronic properties. Au NPs of various shapes and sizes can be synthesized by adopting different methods (physical, chemical and biological). The Au nanospheres were primary accomplishment in the field of Au NPs, while they were not perfectly spherical. Surfactant, polymers, pharmaceuticals, DNA, RNA, proteins and oligonucleotides are some of the

functionalizing agents with which Au NPs can be conjugated [23]. They have broad applications in the fields of environment sensing, electronics, biomedicine and fine chemical synthesis. Because of their multivalency, gold nanoparticles can shield unstable medicines or inadequately soluble imaging contrast agents and facilitate their effective delivery to else inapproachable regions of the body [24]. Optical property of gold nanoparticles helps us to construct optical biosensors such as optical biosensor, electrochemical biosensor and piezoelectric biosensor [25].

The objective of my study is to describe the green synthesis of gold nanoparticles using plants and their biological applications.

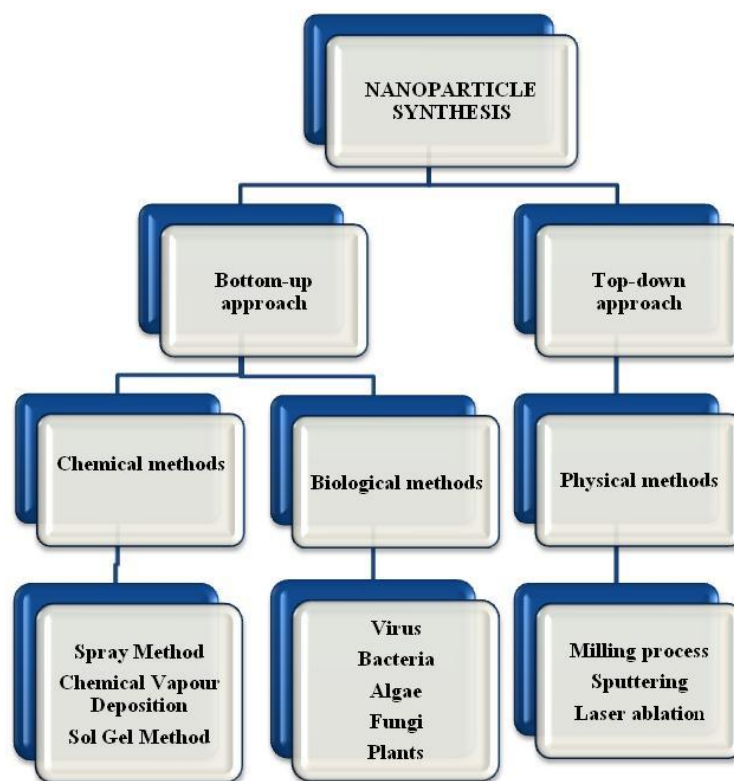
## 2. TECHNIQUES FOR GOLD NANOPARTICLE SYNTHESIS:- There are different methods for Au NPs synthesis as shown in [Figure 2](#).

### 2.1. Top-Down and Bottom-Up Approach:-

Au NPs can be synthesized by two techniques:- top-down approach and bottom-up approach. In top-down approach, a suitable starting molecule is reduced in size, better known as 'modules'. Top-down approach include different physical methods for synthesizing nanoparticles [26]. Different physical methods for nanoparticle synthesis are, molecular beam emphasis, sputter deposition [27], laser desorption, diffusion flame synthesis, milling method, laser ablation [28] and lithographic techniques [29] etc.

In bottom-up approach, the nanoparticles are initially prepared and latterly assembled into final materials by chemicals or biological procedures of synthesis. The bottom-up technique offers the advantage of improving the likelihood of creating nanoparticles with fewer flaws and more uniform chemical compositions.

Bottom-up approach includes chemical and biological methods to synthesize gold nanoparticles. Different chemical methods for synthesizing nanoparticles are electro deposition, sol-gel process [30], chemical vapour deposition soft chemical method, Langmuir Blodgett method [31], co-precipitation method [32] and wet chemical method [33,34]. Biological methods involve plants and microorganisms such as algae, virus, bacteria, fungi etc.



**Figure 2. Different Methods of Nanoparticle Synthesis**

## 2.2. Why we Choose Biological Method for Gold NPs Synthesis:-

In physical and chemical methods, there is a problem about the control of crystal growth, size and size distribution, stability and aggregation of NPs. Due to different drawbacks of chemical [35] and physical methods [36], biological approach for synthesizing NPs have been increasing [37,38,39].

Biological methods are based on green technology aimed at overcoming negative effects on environment. Biological approach utilize biological materials as reducing bioactive agents, isolated from microorganisms and plants that are eco-friendly, prevent pollution and no production of waste, effective synthesis and additional energy saving and its economical production of NPs is also very important [40].

In the biological approach bacteria [41], yeasts [42], [43], fungi, algae [44] and plants [45], [46] etc. may be used for producing green NPs at high scale because they contain reducing and capping agent. In biological methods there is use of non-toxic capping agent, less hazardous reducing agent, and environment friendly solvent, no need of high temperature calcination for the

production of final product, non-toxic organic solvents are used, methods that used plants can be suitably settled for large scale production of nano-particles [47].

### 2.3. Why Plants are Best Agents:-

From all biological entities plants are the best agent because of their diversity, sustainability, and their waste are also eco-friendly. By using plants it is more easy to maintain cell culture and can also be appropriately settled up for large scale production of nanoparticles [48]. Plants can reduce metal ions faster than fungi or bacteria. The plant mediated nanoparticles are more diverse in size and shape comparatively to those produced by other organisms [49].

Biomolecules of plant extracts can be used in single step synthesis of NPs by reducing metal ions to NPs [50,51]. Plants extracts act both as reducing and capping agents for synthesizing nanoparticle. The biomolecules like vitamins, proteins, alkaloids, polysaccharides, amino acids, and organic acids such as citrates that are present in plant extract help in bioreduction of MNPs [52]. We can faster synthesis rate by using plant extracts of high production capability and increasing the reaction temperature. We can also control particle size and shape by changing the plant type or temperature and composition of the reaction mixture [53].

## 3. GENERAL MECHANISM FOR GREEN SYNTHESIS OF NANOPARTICLES FROM PLANTS:-

There are many researches about synthesis of nanoparticles from plants. The general mechanism for synthesis of nanoparticles from plants is presented in [Figure 3](#).

For synthesis of nanoparticles there are three steps: activation, growth and termination.

**Activation:-**Firstly plant parts (fruits, leaves, stems, roots) are washed with distilled water and small pieces are boiled to make the extraction and then mixed with solution of metal ion. Plant extracts comprise bioactive agents such as phenolic acids, sugars, polyphenols, terpenoids, proteins, alkaloids, etc., that firstly reduce the metallic ions and then stabilize them. The metal ions are reduced by biomolecules present in plant extract. The metal salts are converted to metal nanoparticles by oxidation-reduction mechanisms. Therefore, the metals are converted to nanoscale zero-valent metallic particles [54,55].

**Growth phase:-**After the formation of nanoscale metallic particles nucleation takes place. The separated metal atoms congregate to form metal NPs and again biological reduction of metal



ions takes place. Along with the process of nucleation, nanoparticles combine to form divergent shapes like rods, cubes, triangles, pentagons, spheres [56,57].

**Termination phase:**-NPs are stabilized and capped by plant metabolites. Reduction of metal ion is indicated by change in culture colour [16,58]. In this final step nanoparticles obtain their optimistic and stable morphologies.

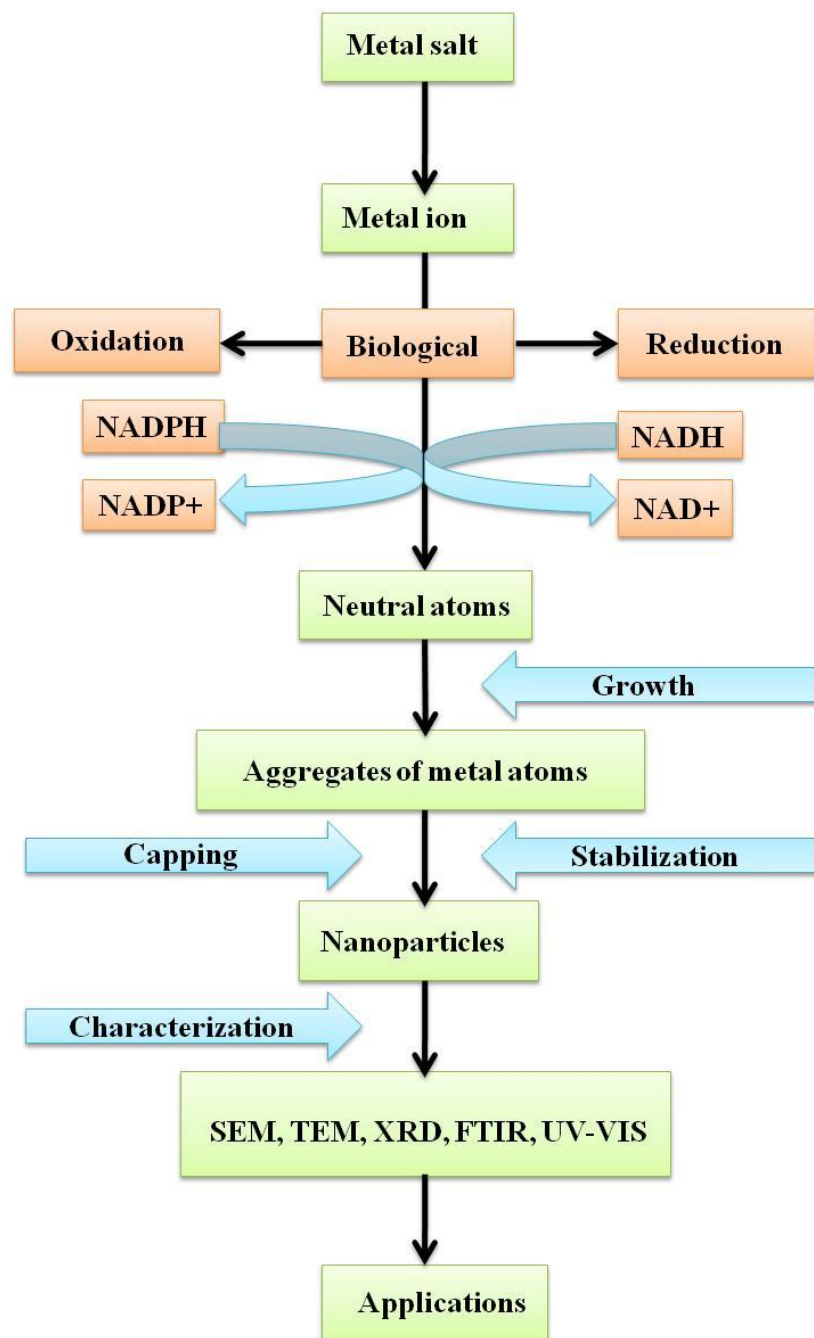


Figure 3. Mechanism for Synthesis of Nanoparticles from Plants



Different plant parts like fruits, leaves, stems, roots and their extracts have been used for different metal nanoparticles synthesis. Many exemplifications are given in **Table 2**.

**Table 2. Synthesis of Different Nanoparticles Using Various Plant Parts and their Activities**

S.No.	Plants	Plant part used	Nanoparticles	Activity of nanoparticle	References
1	<i>Acalypha indica</i>	Leaves	Ag	Antimicrobial activity	[59]
2	<i>Amaranthus dubius</i>	Leaves	Fe	Photocatalytic and Antioxidant activity	[60]
3	<i>Asparagus adscendens</i>	Root and leaves	Cu	Antimicrobial activity	[61]
4	<i>Berberis asiatica</i>	Roots	Ag	Antibacterial activity	[62]
5	<i>Catharanthus roseus</i>	Leaves	Ag	Antiplasmodial activity	[63]
6	<i>Celastrus paniculatus</i>	Leaves	Cu	Photocatalytic and antifungal activity	[64]
7	<i>Cleome viscosa</i>	Fruit	Ag	Antibacterial and anticancer Activity	[65]
8	<i>Coleus aromaticus</i>	Leaves	Ag	Bactericidal activity	[66]
9	<i>Cuminum cyminum</i>	Seed	Cu	Antimicrobial activity	[67]
10	<i>Desmodium gangeticum</i>	Root	Cu	Antimicrobial activity	[68]
11	<i>Diospyros paniculata</i>	Root	Ag	Antimicrobial activity	[69]
12	<i>Euphorbia esula</i>	Leaves	Cu	Catalytic activity	[54]
13	<i>Glycyrrhiza glabra</i>	Root	Ag	Anti-ulcer activity	[70]
14	<i>Lagenaria Siceraria</i>	Leaves	Fe-oxide	Antimicrobial Activity	[71]
15	<i>Lantana camara</i>	Fruit	Ag	Antimicrobial activity	[72]
16	<i>Lawsonia inermis</i>	Leaves	Fe	Antibacterial Activity	[73]
17	<i>Magnolia kobus</i>	Leaves	Cu	Antibacterial activity	[74]
18	<i>Malus domestica</i>	Fruit	Ag	Antimicrobial activity	[75]
19	<i>Moringa oleifera</i>	Seed	Fe	Antibacterial activity	[76]
20	<i>Persea americana</i>	Seeds	Cu	Antimicrobial and	[77]

				antioxidant	
21	<i>Piper nigrum</i>	Seed	SnO <sub>2</sub>	Antitumor activity	[78]
22	<i>Punica granatum</i>	Seed	Fe <sub>2</sub> O <sub>3</sub>	Photocatalytic activity	[79]
23	<i>Rheum palmatum</i>	Root	Ag	Antibacterial activity	[80]
24	<i>Rheum palmatum</i>	Root	CuO <sub>2</sub>	Heterogeneous catalytic activity	[81]
25	<i>Tradescantia spathacea</i>	Leaves	SnO <sub>2</sub>	Photoantioxidant activity	[82]
26	<i>Trigonella foenum-graecum</i>	Seed	Ag and Fe-oxide	Antibacterial and Antioxidant Activities	[83]

#### 4. GREEN SYNTHESIS OF GOLD NANOPARTICLES USING VARIOUS EXTRACTS OF DIFFERENT PLANTS:-

##### 4.1. By Using Leaf Extract:-

Au NPs can be synthesized by the use of leaf extract of different plants. Leaf extract of *Buhinia purpurea* and HAuCl<sub>4</sub> used to prepare gold nanoparticles. Synthesized NPs has excellent anticancer, oxidant, catalytic property. These nanoparticles act as good catalyst for reduction reaction of methylene blue and rhodamine B by NaBH<sub>4</sub>. Synthesized gold nanoparticles have anticancer activity on lung carcinoma cell line [84].

Au NPs of average size 15.3 nm has also been synthesized using leaf extract of *Cacumen platycladi*. Synthesized NPs were of diverse morphology such as sphere, triangles. In this synthesis flavonoids and reducing sugars act both as reducing and protecting agent. Author studied the effect of temperature and pH on size of nanoparticles. Size of NPs decrease with increasing temperature and pH [85].

*Hibiscus-rosa-sinensis*'s leaf extract has also been used to synthesize the gold NPs of different shapes and sizes by varying the ratio of metal salts and extract. From FTIR it was confirmed that amine group was responsible for gold NPs stabilization [86].

Au NPs has also been synthesized using *Cerasus serrulata* leaf extract [87]. The synthesized NPs has size in range from 5 to 25 nm and spherical in shape. *C. serrulata* contain five main compounds which help in the process of reduction. But the major groups were Butylhydroxytoluene, Hydrocoumarin and coumarin; from these coumarin is best which act as

reducing and capping agent for Au NPs synthesis. The synthesized NPs show antibacterial activity against *Escherichia coli* a gram negative bacterium.

*Magnolia kobus* has also been used for synthesizing Au NPs [88]. Nanoplates of width 250-300 nm and thickness of 5-7 nm were formed using 5% leaf broth at 25°C and the impact of concentration and temperature on the synthesis rate and size/shape of gold NPs were studied. It was observed that rate of synthesis increase with the temperature and size of NPs decrease with increasing concentration of leaf extract. From FTIR it was confirmed that Au NPs were surrounded by some proteins and metabolites like terpenoids with numerous functional groups such as alcohols, aldehydes, amines, carboxylic acids and ketones.

*Murraya koenigii* can also be used for the conflation of gold nanoparticle. Varying concentration of leaf extracts of *M. koenigii* was added to HAuCl<sub>4</sub>.3H<sub>2</sub>O to obtain different colloids such as g1, g2, g3, g4 and g5 of different shape and size. Average shape and size of Au NPs prepared was spherical and 20nm respectively. FTIR measurement shows that biomolecules responsible for capping and efficient stabilization in *M. koenigii* leaf are polyphenols, flavonoids and alkaloids [89].

#### 4.2. By Using Root Extract:-

Au NPS of spherical shape and 10-15nm in diameter have been prepared using *Glycyrrhiza uralensis* root extract. The reduction of methylene blue with *Glycyrrhiza* root extract is catalysed by Au NPs that have been prepared. Catalytic activity was confirmed by the decrease in absorbance value of methylene blue in UV-Vis spectra. The in vitro cytotoxicity of Au NPs was appraised by the colour change from purple to yellow at 517nm. These Au NPs are toxic toward murine macrophage and non-toxic toward breast cancer cell lines [90].

Use of *Lanthana camara* Linn root extract has also been reported for synthesis of Au NPs of average size 11-32nm with spherical morphology. It was reported that N-H and O-H functional groups are responsible for reduction of Au ions to Au NPs and flavonoids for capping and stabilization of Au NPs. The *L. camara* Au NPs have dose dependent significant inhibitory effect on breast cancer cell line (MDA-MB-231) and normal cell line (Vero) [91].

Au NPs have also been synthesized using root extract of *Morinda citrifolia*. The synthesized Au NPs were 12.17-38-26 nm in size. From FTIR spectrum, it was reported that proteins present in root extract act as reducing and capping agents [92].

Root extract of *Trianthema decandra* has also been used for the synthesis of Au NPs. Synthesized NPs were of different shapes such as spherical, triangular, hexagonal and cubic with size ranging from 33.7 to 99.3nm. Triangular nanoparticles show excellent antibacterial

activity against *Y. enterocolitica*, *S. faecalis*, *E. coli*, *P. vulgaris* and *S. aureus*. Hence these synthesized Au NPs can be used for treatment of human infection due to microorganisms [93].

Au NPs with various shapes such as triangular, spherical and hexagonal with average size 24.7nm have also been prepared using root extract of *Arctium lappa*. The author reported that synthesized NPs have significant catalytic role for detoxification of common dye Rhodamine B (RhB). Synthesized Au NPs have significant catalytic role for the degradation of 4-Nitrophenol using NaBH<sub>4</sub> [94].

#### 4.3. By Using Seed Extract:-

Au NPs synthesized from seed extract of *Abelmoschus esculentus* were of various sizes and the reason was that the nucleation and aggregation occur consecutively. FTIR shows that –OH functional group in the extract act as capping agent in NPs synthesis. The synthesized Au NPs have antifungal activity against *P. graminis*, *A. flavus*, *A. niger* and *C. albicans*. Hence these NPs have great potential to be used for drug preparation against fungal diseases [95].

Seed extract of *Benincasa hispida* has also been used for Au NPs synthesis. Different colloids were prepared using different concentrations of extract to solution of chloroauric acid. The reduction process was fast when 35ml of extract was used against 30ml solution of chloroauric acid. The produced particles were approximately spherical in form, according to TEM image. The polyol present in seed extract help in reducing chloroauric acid and –COOH group present in protein aid in stability of Au NPs [96].

Polydispersed Au NPs can be synthesized using *Cuminum cyminum* seed extract. The author reported the effect of pH and temperature on morphology the NPs. Increase in temperature and pH result in the production of spherical Au NPs and triangular nano-plates obtained at lower temperature and pH [97].

The Au NPs synthesized from seed extract of *Terminalia chebula* has significant antimicrobial activity. Antimicrobial properties were studied using *E. coli* and *S. aureus* but TC-Au NPs shows antimicrobial activity against *S. aureus* [98].

Au NPs of spherical shape with average size 15.2 nm has been synthesized using *Elettaria cardamomum* seed extract. Au NPs (B<sub>3</sub>) synthesized using 10ml of extract against 30ml solution of HAuCl<sub>4</sub> has significant antioxidant activity against DPPH, NO and OH. This radical scavenging activity enhance with increasing concentration of B<sub>3</sub> Au NP. These NPs has effective antibacterial activity against *S. aureus*, *E. coli* and *P. aeruginosa*. Due to antibacterial and antimicrobial activity, Au NPs (B<sub>3</sub>) has significant application in pharmaceutical. The synthesized

Au NPs has cytotoxicity aid against HeL (Human erythro leukemia) a cell lines, so it can be used for human cancer therapy [99].

#### 4.4. By Using Fruit Extract:-

Polydispersed Au NPs with different shapes and average size 20nm has been synthesized using fruit extract of *Hovenia dulcis*. The synthesized Au NPs have maximum antioxidant activity. These Au NPs have radical scavenging activities for DPPH and hydrogen peroxide. These Au NPs are likewise antimicrobial against *E. coli* and *S. aureus* bacteria. [100].

*Citrus macroptera*'s fruit extract has also been used for Au NPs synthesis. It contains citric acid and ascorbic acid that aid in the reduction of Au<sup>3+</sup> ions to Au particles, as well as carboxylic, amide and hydroxylic groups, which help to stabilize Au NPs. The synthesized Au NPs have antibiofilm activity. These Au NPs prevent biofilm growth of *Pseudomonas aeruginosa* by inhibiting the pyocyanin formation that is responsible for biofilm formation. These Au NPs also has cytotoxic effect against three human cancer cell lines, MDA-MB 468 (human breast cancer cell), A549 (adenocarcinogenic human alveolar basal epithial cell) and HepG2 (human liver cancer cell line) [101].

Au NPs synthesized from *Terminalia arjuna* fruit extract are predominantly spherical in morphology with 25nm average size. Tannin, saponins, terpenoid, glycosides, flavonoids and polyphenolic compounds of the fruit extract of T. arjuna act as reducing and capping agents for Au NPs's synthesis. TA-Au NPs helps to improve seed germination and plant vegetative growth of seedling of *Gloriosa superba* [102].

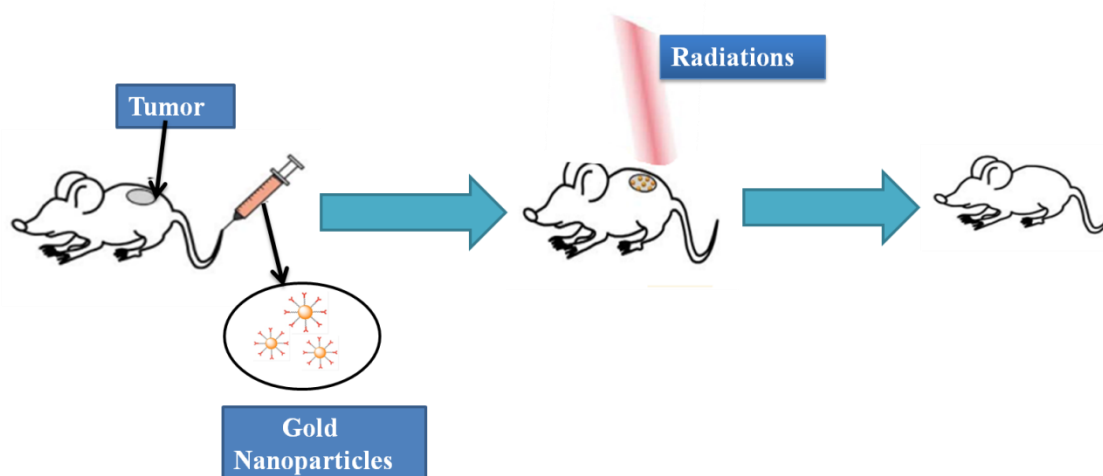
Au NPs of sizes 5 to 10nm can be synthesized using *Lycopersicon esculentum* (Tomato) fruit extract. Tomato juice contains citric acid and ascorbic acid that act as reducing agent. Tomato extract do not act as capping agent, so in this synthesis sodium dodecyl sulfate (SDS) act as capping agent. Methyl parathion can be estimated and detected by using LE-Au NPs in the presence of SDS, hence these Au NPs act as colorimetric sensor [103].

Polydisperse and non-spherical Au NPs can be synthesized using *Mimusops elengi* fruit extract. Antibacterial activity against gram negative bacteria (*E. coli*) and gram positive bacteria (*S. aureus*) and antifungal activity against *Aspergillus niger* and *Aspergillus tubingensis* of synthesized Au NPs has also been reported. [104].

## 5. BIOLOGICAL APPLICATIONS OF GOLD NANOPARTICLES:-

### 5.1. Cancer Treatment Using Photothermal Therapy:-

Cancer is unregulated growth of cells which may spread to other parts of body. Due to unique property of absorption and scattering of electromagnetic radiation, high biocompatibility and high photothermal efficiency, Au NPs are used for the treatment of cancer using photothermal method [105].

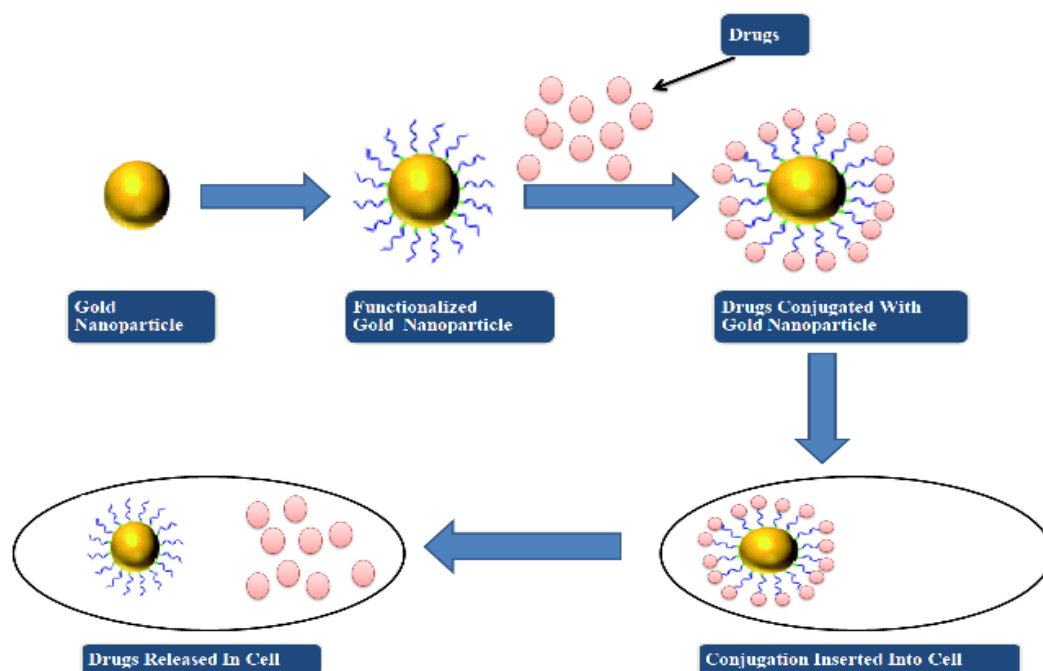


**Figure 4. Schematic Diagram Showing Photothermal Therapy Using Au NPs**

Biological tissue and water rarely absorb any radiation, so Au NPs with SPR frequency in near-infrared region are best source for bioimaging in locating the tumors. Surface modification of NPs are done to provide protection against aggregation, for specific interactions with cells using tumor specific agents, as well as targeted transport and accumulation in desired organs [106]. Au NPs engulfed in tumor cell absorb radiations and convert them into heat causing hyperthermia of tumor cells leading to cell death [107,108].

### 5.2. Drug Delivery:-

Au NPs can also be used as vehicle for drug delivery in treatment of diseases. For drug delivery Au NPs's surfaces are modified with cationic polymers or reactive functional groups (e.g. thiol, carboxyl and amine) [109]. Drugs are adsorbed on the surface of Au NP either covalently or noncovalently. Au NP with drug entered into cell either by gene gun or by natural uptake by cell. Inside the cell Au NP release the drug, this release may be due to natural cell functioning or induced by different methods (e.g. photothermal) [110]. There are various experiments that have proven the functioning of Au NPs in drug delivery [111,112].



**Figure 5. Schematic Diagram for Drug Delivery Using Gold Nanoparticle**

### 5.3. Biosensors:-

Generally sensors are used to specify the presence of analytes and read outs are used for analyte concentration determination. There is change in optical property of gold nanoparticle if optical read out is used. Cysteine-linked core-satellite Au NPs has been used for detection of  $\text{Co}^{2+}$  spiked in diluted human serum [113]. Au NP's biosensor has been developed for the detection of *E. coli* O157: H7. The bacteria were detected by measuring the electrochemical signal of Au NPs [114].

### 5.4. Enhancement in Plant Growth:-

Au NPs helps in enhancing plant growth and seed yield. In *Brassica juncea* Au NPs increased permeability of seed capsule, more light absorbance by chlorophyll thus fastening the photochemical reaction and increase in number of leaves [115]. Seed yield and growth of *Arabidopsis thaliana* also enhanced by Au NPs [116]. Au NPs synthesized using fruit extract of *Terminalia arjuna*, have significant effect on inducing seed germination and growth of *Gloriosa superba* plant [102].

## 6. CONCLUSIONS:-



The review article concluded that, nanotechnology deals with study of particle at nanoscale and great application in real world. Due to small size nanoparticles have improved chemical, physical, and biological properties. Green nanotechnology that works on principles of green chemistry provides green, safe and sustainable method for nanosynthesis. There are different green methods to prepare gold nanoparticles; the present study involves synthesizes gold nanoparticles using plants extracts because of their diversity, sustainability and eco-friendly nature. It is concluded that gold nanoparticles synthesized using plants extract such as leaf, root, seed and fruit extracts have antioxidant and antimicrobial activity. Due to their SPR frequency, biocompatibility and tunable property gold nanoparticles has great application in cancer treatment, biosensor, drug delivery and enhancement of plant growth.

## 7. REFERENCES:-

- (1) Thrall, J. H. Nanotechnology and Medicine. *Radiology* **2004**, *230* (2), 315–318.
- (2) Robinson, P. R.; Hsu, C. S. Introduction to Petroleum Technology. *Springer Handbooks* **2017**, *PartF1*, 1–83..
- (3) Nasrollahzadeh, M.; Sajadi, S. M.; Sajjadi, M.; Issaabadi, Z. *An Introduction to Nanotechnology*, 1st ed.; Elsevier Ltd., 2019; Vol. 28.
- (4) Wang, Z.; Hu, T.; Liang, R.; Wei, M. Application of Zero-Dimensional Nanomaterials in Biosensing. *Front. Chem.* **2020**, *8*, 1–19.
- (5) Jin, T.; Han, Q.; Wang, Y.; Jiao, L. 1D Nanomaterials: Design, Synthesis, and Applications in Sodium–Ion Batteries. *Small* **2018**, *14* (2), 1–26.
- (6) Garnett, E.; Mai, L.; Yang, P. Introduction: 1D Nanomaterials/Nanowires. *Chem. Rev.* **2019**, *119* (15), 8955–8957.
- (7) Chen, C.; Fan, Y.; Gu, J.; Wu, L.; Passerini, S.; Mai, L. One-Dimensional Nanomaterials for Energy Storage. *J. Phys. D. Appl. Phys.* **2018**, *51* (11).
- (8) Zhuang, X.; Mai, Y.; Wu, D.; Zhang, F.; Feng, X. Two-Dimensional Soft Nanomaterials: A Fascinating World of Materials. *Adv. Mater.* **2015**, *27* (3), 403–427..
- (9) Ashton, T. S.; Moore, A. L. Three-Dimensional Foam-like Hexagonal Boron Nitride Nanomaterials via Atmospheric Pressure Chemical Vapor Deposition. *J. Mater. Sci.* **2015**, *50* (18), 6220–6226.
- (10) Elakkiya, R.; Maduraiveeran, G. A Three-Dimensional Nickel-Cobalt Oxide Nanomaterial as an Enzyme-Mimetic Electrocatalyst for the Glucose and Lactic Acid Oxidation Reaction. *New J. Chem.* **2019**, *43* (37), 14756–14762.
- (11) Sridhar, V.; Kim, H. J.; Jung, J. H.; Lee, C.; Park, S.; & Oh, I. K. Defect-Engineered Three-Dimensional Graphene- Nanotube - Palladium Nanostructures with Ultrahigh Capacitance. *ACS Nano* **2012**, *6* (12), 10562–10570.
- (12) Bogunia-Kubik, K.; Sugisaka, M. From Molecular Biology to Nanotechnology and Nanomedicine. *BioSystems* **2002**, *65* (2–3), 123–138..
- (13) Christian, P.; Von Der Kammer, F.; Baalousha, M.; Hofmann, T. Nanoparticles: Structure, Properties, Preparation and Behaviour in Environmental Media. *Ecotoxicology* **2008**, *17* (5), 326–343..
- (14) Hussain, I.; Singh, N. B.; Singh, A.; Singh, H.; Singh, S. C. Green Synthesis of Nanoparticles and Its Potential Application. *Biotechnol. Lett.* **2016**, *38* (4), 545–560..

- (15) Tripathi, D.; Modi, A.; Narayan, G.; Rai, S. P. Green and Cost Effective Synthesis of Silver Nanoparticles from Endangered Medicinal Plant *Withania Coagulans* and Their Potential Biomedical Properties. *Mater. Sci. Eng. C* **2019**, *100*, 152–164.
- (16) Sadeghi, B.; Gholamhoseinpoor, F. A Study on the Stability and Green Synthesis of Silver Nanoparticles Using *Ziziphora Tenuior* (Zt) Extract at Room Temperature. *Spectrochim. Acta - Part A Mol. Biomol. Spectrosc.* **2015**, *134*, 310–315.
- (17) Fischer, H. C.; Chan, W. C. Nanotoxicity: The Growing Need for in Vivo Study. *Curr. Opin. Biotechnol.* **2007**, *18* (6), 565–571..
- (18) Varma, R. S.; States, U.; Protection, E. Journey on Greener Pathways: From the Use of Alternate Energy Inputs and Benign Reaction Media to Sustainable Applications of Nano-Catalysts in Synthesis and Environmental Remediation. *Green Chem.* **2014**, *16* (4), 2027–2041.
- (19) Backx, B. P. Green Nanotechnology: Only the Final Product That Matters? *Nat. Prod. Res.* **2020**, *0* (0), 1–3.
- (20) Assistant, M.; Reddy, R.; Assistant, N.; Lemoor, T. Research Project On Green Nanotechnology Usage In Automobiles. **2016**, No. 4, 5144–5156.
- (21) Miklicanin, M. M. and E. O.-. Towards Green Nanotechnology: Maximizing Benefits and Minimizing Harm Mirjana. *CMBEBIH* **2017**, 164–170.
- (22) Jahangirian, H.; Lemraski, E. G.; Webster, T. J.; Rafiee-Moghaddam, R.; Abdollahi, Y. A Review of Drug Delivery Systems Based on Nanotechnology and Green Chemistry: Green Nanomedicine. *Int. J. Nanomedicine* **2017**, *12*, 2957–2978.
- (23) Sun, L.; Liu, D.; Wang, Z. Functional Gold Nanoparticle - Peptide Complexes as Cell-Targeting Agents. *Langmuir* **2008**, *24* (18), 10293–10297.
- (24) Dreaden, E. C.; Alkilany, A. M.; Huang, X.; Murphy, C. J.; El-Sayed, M. A. The Golden Age: Gold Nanoparticles for Biomedicine. *Chem. Soc. Rev.* **2012**, *41* (7), 2740–2779.
- (25) Li, Y.; Schluesenerb, H.J.; Xu, S. Gold Nanoparticle Based Biosensors. *New Dev. Gold Nanomater. Res.* **2010**, *43* (1), 29–41.
- (26) Ghorbani, H. R. A Review of Methods for Synthesis of Al Nanoparticles. *Orient. J. Chem.* **2014**, *30* (4), 1941–1949.
- (27) Chung, M. W.; Cha, I. Y.; Ha, M. G.; Na, Y.; Hwang, J.; Ham, H. C.; Kim, H. J.; Henkensmeier, D.; Yoo, S. J.; Kim, J. Y.; Lee, S. Y.; Park, H. S.; Jang, J. H. Enhanced CO<sub>2</sub> Reduction Activity of Polyethylene Glycol-Modified Au Nanoparticles Prepared via Liquid Medium Sputtering. *Appl. Catal. B Environ.* **2018**, *237*, 673–680.
- (28) Kimura, Y.; Takata, H.; Terazima, M.; Ogawa, T.; Isoda, S. Preparation of Gold Nanoparticles

- by the Laser Ablation in Room-Temperature Ionic Liquids. *Chem. Lett.* **2007**, *36* (9), 1130–1131.
- (29) Corbierre, M. K.; Beerens, J.; Lennox, R. B. Gold Nanoparticles Generated by Electron Beam Lithography of Gold(I)-Thiolate Thin Films. *Chem. Mater.* **2005**, *17*, 5774–5779.
- (30) Kobayashi, Y.; Correa-duarte, M. A.; Liz-marza, L. M. Sol - Gel Processing of Silica-Coated Gold Nanoparticles. *Langmuir* **2001**, *17* (20), 6375–6379.
- (31) Heriot, S. Y.; Zhang, H. L.; Evans, S. D.; Richardson, T. H. Multilayers of 4-Methylbenzenethiol Functionalized Gold Nanoparticles Fabricated by Langmuir-Blodgett and Langmuir-Schaefer Deposition. *Colloids Surfaces A Physicochem. Eng. Asp.* **2006**, *278* (1–3), 98–105.
- (32) Wang, C. T.; Chen, H. Y.; Chen, Y. C. Gold/Vanadium-Tin Oxide Nanocomposites Prepared by Co-Precipitation Method for Carbon Monoxide Gas Sensors. *Sensors Actuators, B Chem.* **2013**, *176*, 945–951.
- (33) Palui, G.; Ray, S.; Banerjee, A. Synthesis of Multiple Shaped Gold Nanoparticles Using Wet Chemical Method by Different Dendritic Peptides at Room Temperature. *J. Mater. Chem.* **2009**, *19* (21), 3457–3468.
- (34) Jana, N. R.; Gearheart, L.; Murphy, C. J. Wet Chemical Synthesis of High Aspect Ratio Cylindrical Gold Nanorods. *J. Phys. Chem. B* **2001**, *105* (19), 4065–4067.
- (35) Li, X.; Xu, H.; Chen, Z. S.; Chen, G. Biosynthesis of Nanoparticles by Microorganisms and Their Applications. *J. Nanomater.* **2011**, *2011*, 1–16.
- (36) Thakkar, K. N.; Mhatre, S. S.; Parikh, R. Y. Biological Synthesis of Metallic Nanoparticles. *Nanomedicine Nanotechnology, Biol. Med.* **2010**, *6* (2), 257–262.
- (37) Mukherjee, P.; Ahmad, A.; Mandal, D.; Senapati, S.; Sainkar, S. R.; Khan, M. I.; Parishcha, R.; Ajaykumar, P. V.; Alam, M.; Kumar, R.; Sastry, M. Fungus-Mediated Synthesis of Silver Nanoparticles and Their Immobilization in the Mycelial Matrix: A Novel Biological Approach to Nanoparticle Synthesis. *Nano Lett.* **2001**, *1* (10), 515–519.
- (38) Mohanpuria, P.; Rana, N. K.; Yadav, S. K. Biosynthesis of Nanoparticles: Technological Concepts and Future Applications. *J. Nanoparticle Res.* **2008**, *10* (3), 507–517.
- (39) Dhillon, G. S.; Brar, S. K.; Kaur, S.; Verma, M. Green Approach for Nanoparticle Biosynthesis by Fungi: Current Trends and Applications. **2012**, *32* (October 2010), 49–73.
- (40) Salem, S. S.; Fouda, A. Green Synthesis of Metallic Nanoparticles and Their Prospective Biotechnological Applications: An Overview. *Biol. Trace Elem. Res.* **2021**, *199* (1), 344–370.
- (41) He, S.; Guo, Z.; Zhang, Y.; Zhang, S.; Wang, J.; Gu, N. Biosynthesis of Gold Nanoparticles

- Using the Bacteria *Rhodospseudomonas Capsulata*. *Mater. Lett.* **2007**, *61* (18), 3984–3987.
- (42) Zhang, X.; Qu, Y.; Shen, W.; Wang, J.; Li, H.; Zhang, Z.; Li, S.; Zhou, J. Biogenic Synthesis of Gold Nanoparticles by Yeast *Magnusiomyces Ingens* LH-F1 for Catalytic Reduction of Nitrophenols. *Colloids Surfaces A Physicochem. Eng. Asp.* **2016**, *497*, 280–285.
- (43) Mourato, A.; Gadanho, M.; Lino, A. R.; Tenreiro, R. Biosynthesis of Crystalline Silver and Gold Nanoparticles by Extremophilic Yeasts. *Bioinorg. Chem. Appl.* **2011**, *2011*.
- (44) Singh, M.; Kalaivani, R.; Manikandan, S.; Sangeetha, N.; Kumaraguru, A. K. Facile Green Synthesis of Variable Metallic Gold Nanoparticle Using *Padina Gymnospora*, a Brown Marine Macroalga. *Appl. Nanosci.* **2013**, *3* (2), 145–151.
- (45) Unal, I. S.; Demirbas, A.; Onal, I.; Ildiz, N.; Ocsoy, I. One Step Preparation of Stable Gold Nanoparticle Using Red Cabbage Extracts under UV Light and Its Catalytic Activity. *J. Photochem. Photobiol. B Biol.* **2020**, *204*, 111800.
- (46) Sadeghi, B.; Mohammadzadeh, M.; Babakhani, B. Green Synthesis of Gold Nanoparticles Using *Stevia Rebadiauna* Leaf Extracts: Characterization and Their Stability. *J. Photochem. Photobiol. B Biol.* **2015**, *148*, 101–106.
- (47) Nasrollahzadeh, M.; Sajadi, S. M. Synthesis and Characterization of Titanium Dioxide Nanoparticles Using *Euphorbia Heteradena* Jaub Root Extract and Evaluation of Their Stability. *Ceram. Int.* **2015**, *41* (10), 14435–14439.
- (48) S. Shiv Shankar, Akhilesh Rai, Absar Ahmad, and M. S. Rapid Synthesis of Au , Ag , and Bimetallic Au Core – Ag Shell Nanoparticles Using Neem ( *Azadirachta Indica* ) Leaf Broth. *J. Colloid Interface Sci.* **2004**, *275* (2), 496–502.
- (49) Iravani, S. Green Synthesis of Metal Nanoparticles Using Plants. *Green Chem.* **2011**, *13* (10), 2638–2650.
- (50) Nasrollahzadeh, M.; Atarod, M.; Sajjadi, M.; Sajadi, S. M.; Issaabadi, Z. *Plant-Mediated Green Synthesis of Nanostructures: Mechanisms, Characterization, and Applications*, 1st ed.; Elsevier Ltd., 2019; Vol. 28.
- (51) Duan, H.; Wang, D.; Li, Y. Green Chemistry for Nanoparticle Synthesis. *Chem. Soc. Rev.* **2015**, *44* (16), 5778–5792.
- (52) Kavitha, K. S.; Baker, S.; Rakshith, D.; Kavitha, H. U.; C, Y. R. H.; Harini, B. P.; Satish, S. Plants as Green Source towards Synthesis of Nanoparticles. **2013**, *2* (6), 66–76.
- (53) Yong Song, J.; Soo Kim, B. Biological Synthesis of Metal Nanoparticles. *Biocatal. Agric. Biotechnol.* **2009**, 399–407.
- (54) Nasrollahzadeh, M.; Sajadi, S. M.; Khalaj, M. Green Synthesis of Copper Nanoparticles Using

Aqueous Extract of the Leaves of Euphorbia Esula L and Their Catalytic Activity for Ligand-Free Ullmann-Coupling Reaction and Reduction of 4-Nitrophenol. *RSC Adv.* **2014**, 4 (88), 47313–47318.

- (55) Malik, P.; Shankar, R.; Malik, V.; Sharma, N.; Mukherjee, T. K. Green Chemistry Based Benign Routes for Nanoparticle Synthesis. *J. Nanoparticles* **2014**, 2014, 1–14.
- (56) Trewyn, B. G.; Nieweg, J. A.; Zhao, Y.; Lin, V. S. Y. Biocompatible Mesoporous Silica Nanoparticles with Different Morphologies for Animal Cell Membrane Penetration. *Chem. Eng. J.* **2008**, 137 (1), 23–29..
- (57) Gao, M.; Sun, L.; Wang, Z.; Zhao, Y. Controlled Synthesis of Ag Nanoparticles with Different Morphologies and Their Antibacterial Properties. *Mater. Sci. Eng. C* **2013**, 33 (1), 397–404.
- (58) Jahan, I.; Erci, F.; Isildak, I. Microwave-Assisted Green Synthesis of Non-Cytotoxic Silver Nanoparticles Using the Aqueous Extract of Rosa Santana (Rose) Petals and Their Antimicrobial Activity. *Anal. Lett.* **2019**, 52 (12), 1860–1873..
- (59) Krishnaraj, C.; Jagan, E. G.; Rajasekar, S.; Selvakumar, P.; Kalaichelvan, P. T.; Mohan, N. Synthesis of Silver Nanoparticles Using Acalypha Indica Leaf Extracts and Its Antibacterial Activity against Water Borne Pathogens. *Colloids Surfaces B Biointerfaces* **2010**, 76 (1), 50–56.
- (60) Harshiny, M.; Iswarya, C. N.; Matheswaran, M. Biogenic Synthesis of Iron Nanoparticles Using Amaranthus Dubius Leaf Extract as a Reducing Agent. *Powder Technol.* **2015**, 286, 744–749. .
- (61) Thakur, S.; Sharma, S.; Thakur, S.; Rai, R. Green Synthesis of Copper Nano-Particles Using Asparagus Adscendens Roxb . Root and Leaf Extract and Their Antimicrobial Activities Green Synthesis of Copper Nano-Particles Using Asparagus Adscendens Roxb . Root and Leaf Extract and Their Antimicrobial Ac. **2018**, 7 (4), 683–694. .
- (62) Dangi, S.; Gupta, A.; Gupta, D. K.; Singh, S.; Parajuli, N. Green Synthesis of Silver Nanoparticles Using Aqueous Root Extract of Berberis Asiatica and Evaluation of Their Antibacterial Activity. *Chem. Data Collect.* **2020**, 28, 100411.
- (63) Ponarulselvam, S.; Panneerselvam, C.; Murugan, K.; Aarthi, N.; Kalimuthu, K.; Thangamani, S. Synthesis of Silver Nanoparticles Using Leaves of Catharanthus Roseus Linn. G. Don and Their Antiplasmodial Activities. *Asian Pac. J. Trop. Biomed.* **2012**, 2 (7), 574–580.
- (64) Mail, S. C.; Dhaka, A.; Githala, Chanda, K.; Trivedi, R. Green Synthesis of Copper Nanoparticles Using Celastrus Paniculatus Willd. Leaf Extract and Their Photocatalytic and Antifungal Properties. *Biotechnol. Reports* **2020**, 27, e00518.



- (65) Lakshmanan, G.; Sathiyaseelan, A.; Kalaichelvan, P. T.; Murugesan, K. Plant-Mediated Synthesis of Silver Nanoparticles Using Fruit Extract of *Cleome Viscosa* L.: Assessment of Their Antibacterial and Anticancer Activity. *Karbala Int. J. Mod. Sci.* **2018**, *4* (1), 61–68.
- (66) Vanaja, M.; Annadurai, G. *Coleus Aromaticus* Leaf Extract Mediated Synthesis of Silver Nanoparticles and Its Bactericidal Activity. *Appl. Nanosci.* **2013**, *3* (3), 217–223.
- (67) Rajesh, K. M.; Ajitha, B.; Reddy, Y. A. K.; Suneetha, Y.; Reddy, P. S.; Ahn, C. W. A Facile Bio-Synthesis of Copper Nanoparticles Using *Cuminum Cyminum* Seed Extract : Antimicrobial Studies. *Adv. Nat. Sci. Nanosci. Nanotechno.* **2018**, *9* (3), 035005.
- (68) Guin, R.; A, S. B.; Kurian, G. A. Synthesis of Copper Oxide Nanoparticles Using *Desmodium Gangeticum* Aqueous Root Extract Innovare. *Int. J. Pharm. Pharm. Sci.* **2015**, *7*, 60–65.
- (69) Rao, N. H.; Lakshmidhevi, N.; Pammi, S. V. N.; Kollu, P.; Ganapaty, S.; Lakshmi, P. Green Synthesis of Silver Nanoparticles Using Methanolic Root Extracts of *Diospyros Paniculata* and Their Antimicrobial Activities. *Mater. Sci. Eng. C.* **2016**, *62*, 553–557.
- (70) Shreelakshmy, V.; Deepa, M. K. Green Synthesis of Silver Nanoparticles from *Glycyrrhiza Glabra* Root Extract for the Treatment of Gastric Ulcer. *J. Dev. Drugs* **2016**, *5* (2), 152.
- (71) Kanagasubbulakshmi, S.; Kadirvelu, K. Green Synthesis of Iron Oxide Nanoparticles Using *Lagenaria Siceraria* and Evaluation of Its Antimicrobial Activity. **2017**, *2* (4), 422–427.
- (72) Shivkumar, P.; Nethradevi, C.; Renganathan, S. Synthesis of Silver Nanoparticles Using *Lanthana camara* Fruit Extract and its Effect on Pathogens. *Asian J. Pharm. Clin. Res. Vol* **2012**, *5* (3), 97–101.
- (73) Naseem, T.; Farrukh, M. A. Antibacterial Activity of Green Synthesis of Iron Nanoparticles Using *Lawsonia Inermis* and *Gardenia Jasminoides* Leaves Extract. **2015**, *2015*, 7.
- (74) Lee, H.; Song, J. Y.; Kim, B. S. Biological Synthesis of Copper Nanoparticles Using *Magnolia Kobus* Leaf Extract and Their Antibacterial Activity. *J. Chem. Technol. Biotechnol.* **2013**, *88* (11), 1971–1977.
- (75) Roy, K.; Sarkar, C. K.; Ghosh, C. K. Green Synthesis of Silver Nanoparticles Using Fruit Extract of *Malus Domestica* and Study of Its Antimicrobial Activity. *Dig. J. Nanomater. Biostructures* **2014**, *9* (3), 1137–1147.
- (76) Katata-seru, L.; Moremedi, T.; Aremu, O. S.; Bahadur, I. Green Synthesis of Iron Nanoparticles Using *Moringa Oleifera* Extracts and Their Applications: Removal of Nitrate from Water and Antibacterial Activity against *Escherichia Coli*. *J. Mol. Liq.* **2018**, *256*, 296–304.
- (77) Rajeshkumar, S.; Rinitha, G. Nanostructural Characterization of Antimicrobial and Antioxidant Copper Nanoparticles Synthesized Using Novel *Persea Americana* Seeds . *OpenNano* **2018**, *3*,



18–27.

- (78) Tammina, S. K.; Mandal, B. K.; Ranjan, S.; Dasgupta, N. Cytotoxicity Study of Piper Nigrum Seed Mediated Synthesized SnO<sub>2</sub> Nanoparticles towards Colorectal (HCT116) and Lung Cancer (A549) Cell Lines. *J. Photochem. Photobiol. B Biol.* **2017**, *166*, 158–168.
- (79) Bibi, I.; Nazar, N.; Ata, S.; Sultan, M.; Ali, A.; Abbas, A. Green Synthesis of Iron Oxide Nanoparticles Using Pomegranate Seeds Extract and Photocatalytic Activity Evaluation for the Degradation of Textile Dye. *J. Mater. Res. Technol.* **2019**, *8* (6), 6115–6124.
- (80) Arokiyaraj, S.; Vincent, S.; Saravanan, M.; Lee, Y.; Oh, Y. K.; Kim, K. H. Green Synthesis of Silver Nanoparticles Using Rheum Palmatum Root Extract and Their Antibacterial Activity against Staphylococcus Aureus and Pseudomonas Aeruginosa. *Artif. Cells, Nanomedicine Biotechnol.* **2017**, *45* (2), 372–379.
- (81) Bordbar, M.; Shari, Z. Green Synthesis of Copper Oxide Nanoparticles / Clinoptilolite Using Rheum Palmatum L . Root Extract : High Catalytic Activity for Reduction of 4-Nitro Phenol , Rhodamine B , and Methylene Blue. *J Sol-Gel Sci Technol* **2017**, *81* (3), 724–733.
- (82) Matussin, S. N.; Harunsani, M. H.; Tan, A. L.; Mohammad, A.; Cho, M. H.; Khan, M. M. Photoantioxidant Studies of the SnO<sub>2</sub> Nanoparticles Fabricated Using Aqueous Leaf Extract of Tradescantia Spathacea. *Solid State Sci.* **2020**, *105*, 106279.
- (83) Deshmukh, A. R.; Gupta, A.; Kim, B. S. Ultrasound Assisted Green Synthesis of Silver and Iron Oxide Nanoparticles Using Fenugreek Seed Extract and Their Enhanced Antibacterial and Antioxidant Activities. *Biomed Res. Int.* **2019**, 1-14.
- (84) Vijayan, R. Anticancer , Antimicrobial , Antioxidant , and Catalytic Activities of Green-Synthesized Silver and Gold Nanoparticles Using Bauhinia Purpurea Leaf Extract. *Bioprocess Biosyst. Eng.* **2019**, *42* (2), 305–319.
- (85) Zhan, G.; Huang, J.; Lin, L. Synthesis of Gold Nanoparticles by Cacumen Platycladi Leaf Extract and Its Simulated Solution : Toward the Plant-Mediated Biosynthetic Mechanism. **2011**, *13* (10), 4957–4968.
- (86) Philip, D. Green Synthesis of Gold and Silver Nanoparticles Using Hibiscus Rosa Sinensis. *Phys. E Low-dimensional Syst. Nanostructures* **2010**, *42* (5), 1417–1424.
- (87) Karthik, R.; Chen, S.; Elangovan, A.; Muthukrishnan, P.; Shanmugam, R.; Lou, B.S. Phyto Mediated Biogenic Synthesis of Gold Nanoparticles Using Cerasus Serru- Lata and Its Utility in Detecting Hydrazine, Microbial Activity and DFT Studies. *J. Colloid Interface Sci.* **2016**, *468*, 163-175
- (88) Song, J. Y.; Jang, H.; Kim, B. S. Biological Synthesis of Gold Nanoparticles Using Magnolia

- Kobus and Diopyros Kaki Leaf Extracts. **2009**, *44*, 1133–1138.
- (89) Philip, D.; Unni, C.; Aromal, S. A.; Vidhu, V. K. Spectrochimica Acta Part A : Molecular and Biomolecular Spectroscopy Murraya Koenigii Leaf-Assisted Rapid Green Synthesis of Silver and Gold Nanoparticles. *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* **2011**, *78*, 899–904.
- (90) Huo, Y.; Singh, P.; Kim, Y. J.; Soshnikova, V.; Markus, J.; Ahn, S.; Castro-aceituno, V.; Chokkalingam, M.; Bae, K.; Yang, D. C.; Huo, Y.; Singh, P.; Kim, Y. J.; Soshnikova, V. Biological Synthesis of Gold and Silver Chloride Nanoparticles by Glycyrrhiza Uralensis and in Vitro Applications. *Artif. Cells, Nanomedicine, Biotechnol.* **2018**, *46* (2), 303–312.
- (91) Rajendiran Ramkumar, Govindasamy Balasubramani, Ramalingam Karthik Raja, Manickam Raja, Raji Govindan, E. K. G. & P. P. Lantana Camara Linn Root Extract-Mediated Gold Nanoparticles and Their in Vitro Antioxidant and Cytotoxic Potentials. *Artif. Cells, Nanomedicine, Biotechnol.* **2017**, *45* (4), 748–757.
- (92) Suman, T. Y.; Rajasree, S. R. R.; Ramkumar, R.; Rajthilak, C.; Perumal, P. Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy The Green Synthesis of Gold Nanoparticles Using an Aqueous Root Extract of Morinda Citrifolia L. *Spectrochim. ACTA PART A Mol. Biomol. Spectrosc.* **2014**, *118*, 11–16.
- (93) Geethalakshmi, R. and Sarada, D. V. L. Gold and Silver Nanoparticles from Trianthema Decandra : Synthesis , Characterization , and Antimicrobial Properties. *Int. J. Nanomedicine* **2012**, *7*, 5375–5384.
- (94) Nguyen, T. T.; Vo, T.; Nguyen, B. N.; Nguyen, D.; Dang, V. Silver and Gold Nanoparticles Biosynthesized by Aqueous Extract of Burdock Root , Arctium Lappa as Antimicrobial Agent and Catalyst for Degradation of Pollutants. *Environ. Sci. Pollut. Res.* **2018**, *25* (34), 34247–34261.
- (95) Jayaseelan, C.; Ramkumar, R.; Abdul, A.; Perumal, P. Green Synthesis of Gold Nanoparticles Using Seed Aqueous Extract of Abelmoschus Esculentus and Its Antifungal Activity. *Ind. Crop. Prod.* **2013**, *45*, 423–429.
- (96) Aromal, S. A.; Philip, D. Benincasa Hispida Seed Mediated Green Synthesis of Gold Nanoparticles and Its Optical Nonlinearity. *Phys. E Low-dimensional Syst. Nanostructures* **2012**, *44*, 1329–1334.
- (97) Sneha, K.; Sathishkumar, M.; Lee, S. Y.; Bae, M. A.; Yun, Y. Biosynthesis of Au Nanoparticles Using Cumin Seed Powder Extract. *J. Nanosci. Nanotechnol.* **2011**, *11*, 1811–1814.
- (98) Kumar, K. M.; Mandal, B. K.; Sinha, M.; Krishnakumar, V. Spectrochimica Acta Part A : Molecular and Biomolecular Spectroscopy Terminalia Chebula Mediated Green and Rapid

- Synthesis of Gold Nanoparticles. *Spectrochim. Acta Part A Mol. Biomol. Spectrosc.* **2012**, *86*, 490–494.
- (99) Rajan, A.; Rajan, A. R.; Philip, D. Elettaria Cardamomum Seed Mediated Rapid Synthesis of Gold Nanoparticles and Its Biological Activities. *OpenNano* **2017**, *2*, 1–8.
- (100) Basavegowda, N.; Idhayadhulla, A.; Lee, Y. R. Phyto-Synthesis of Gold Nanoparticles Using Fruit Extract of Hovenia Dulcis and Their Biological Activities. *Ind. Crop. Prod.* **2014**, *52*, 745–751..
- (101) Majumdar, M.; Biswas, S. C.; Rupasree Choudhury, P. U.; Adhikary, A.; Dijendra Nath Roy, and T. K. M.; Citrus. Synthesis of Gold Nanoparticles Using Citrus Macroptera Fruit Extract : Anti-Biofilm and Anticancer Activity. *Chem. Sel.* **2019**, *4* (19), 5714–5723.
- (102) Gopinath, K.; Gowri, S.; Karthika, V.; Arumugam, A. Green Synthesis of Gold Nanoparticles from Fruit Extract of Terminalia Arjuna , for the Enhanced Seed Germination Activity of Gloriosa Superba. *J. Nanostructure Chem.* **2014**, *4*, 115.
- (103) Barman, G.; Maiti, S.; Laha, J. K. Bio-Fabrication of Gold Nanoparticles Using Aqueous Extract of Red Tomato and Its Use as a Colorimetric Sensor. *Nanoscale Res. Lett.* **2013**, *8* (1), 1–9.
- (104) Tripathy, A.; Behera, M.; Rout, A. S.; Biswal, S. K. Optical , Structural , and Antimicrobial Study of Gold Nanoparticles Synthesized Using an Aqueous Extract of Mimusops Elengi Raw Fruits. *Biointerface Res. Appl. Chem.* **2020**, *10* (6), 7085–7096.
- (105) Sztandera, K.; Gorzkiewicz, M.; Klajnert-Maculewicz, B. Gold Nanoparticles in Cancer Treatment. *Mol. Pharm.* **2019**, *16* (1), 1–23.
- (106) Chinen, A. B.; Guan, C. M.; Ferrer, J. R.; Barnaby, S. N.; Merkel, T. J.; Mirkin, C. A. Nanoparticle Probes for the Detection of Cancer Biomarkers, Cells, and Tissues by Fluorescence. *Chem. Rev.* **2015**, *115* (19), 10530–10574.
- (107) Chen, J.; Ma, Y.; Du, W.; Dai, T.; Wang, Y.; Jiang, W.; Wan, Y.; Wang, Y.; Liang, G.; Wang, G. Furin-Instructed Intracellular Gold Nanoparticle Aggregation for Tumor Photothermal Therapy. *Adv. Funct. Mater.* **2020**, *30* (50), 1–8.
- (108) Hwang, S.; Nam, J.; Jung, S.; Song, J.; Doh, H.; Kim, S. Gold Nanoparticle-Mediated Photothermal Therapy: Current Status and Future Perspective. *Nanomedicine* **2014**, *9* (13), 2003–2022.
- (109) Duncan, B.; Kim, C.; Rotello, V. M. Gold Nanoparticle Platforms as Drug and Biomacromolecule Delivery Systems. *J. Control. Release* **2010**, *148*, 122–127.

- (110) Niikura, K.; Iyo, N.; Matsuo, Y.; Mitomo, H.; Ijro, K. Sub-100 Nm Gold Nanoparticle Vesicles as a Drug Delivery Carrier Enabling Rapid Drug Release upon Light Irradiation. *ACS Appl. Mater. Interfaces* **2013**, 5 (9), 3900–3907.
- (111) Ajnai, G.; Chiu, A.; Kan, T.; Cheng, C. C.; Tsai, T. H.; Chang, J. Trends of Gold Nanoparticle-Based Drug Delivery System in Cancer Therapy. *J. Exp. Clin. Med.* **2014**, 6 (6), 172–178.
- (112) Liang, J.-J.; Zhou, Y.-Y.; Wu, J.; Ding, Y. Gold Nanoparticle-Based Drug Delivery Platform for Antineoplastic Chemotherapy. *Curr. Drug Metab.* **2014**, 15, 620–631.
- (113) Mazur, F.; Liu, L.; Li, H.; Huang, J.; Chandrawati, R. Core-Satellite Gold Nanoparticle Biosensors for Monitoring Cobalt Ions in Biological Samples. *Sensors Actuators, B Chem.* **2018**, 268, 182–187. .
- (114) Wang, Y.; Alocilja, E. C. Gold Nanoparticle-Labeled Biosensor for Rapid and Sensitive Detection of Bacterial Pathogens. *J. Biol. Eng.* **2015**, 9 (1), 1–7.
- (115) Arora, S.; Sharma, P. Gold-Nanoparticle Induced Enhancement in Growth and Seed Yield of Brassica Juncea. *Plant Growth Regul.* **2012**, 66 (3), 303–310. .
- (116) Kumar, V.; Guleria, P.; Kumar, V.; Yadav, S. K. Gold Nanoparticle Exposure Induces Growth and Yield Enhancement in Arabidopsis Thaliana. *Sci. Total Environ.* **2013**, 461, 462–468.